

YMC-TriArt

Versatile hybrid silica based (U)HPLC columns

YMC
EUROPE GMBH
The Selectivity Company



Transfer

Scalable particles:
EASY
UHPLC ↔ HPLC

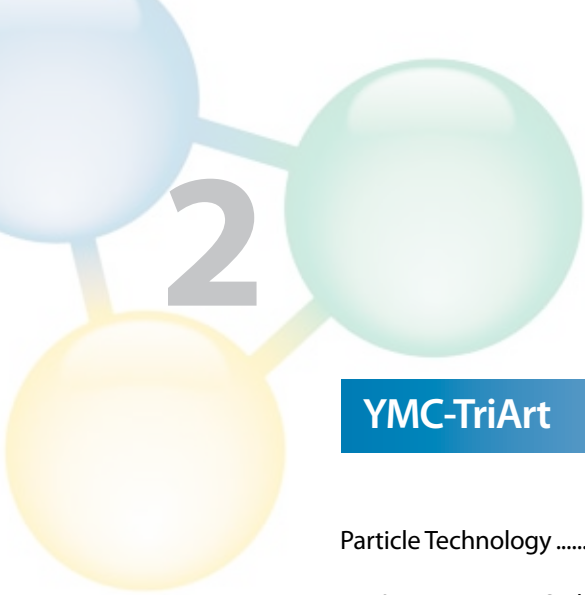
Flexible

YMC-TriArt:
pH 1-12
Temperatures
up to 70°C

Universal

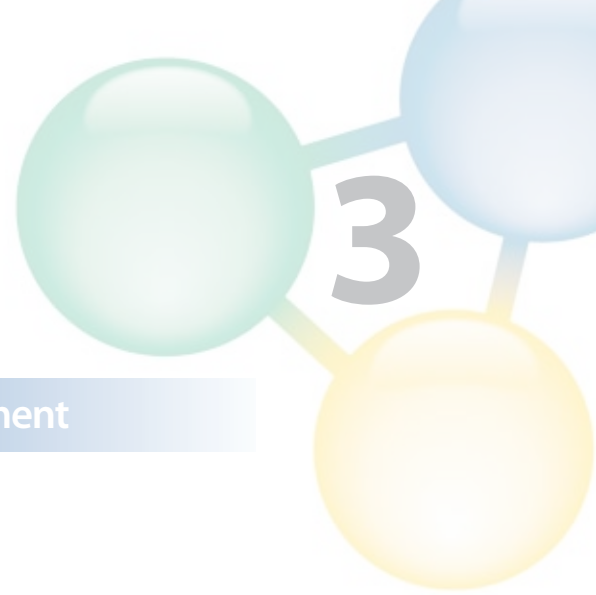
YMC-TriArt
for acidic, basic and
neutral analytes

www.ymc.de

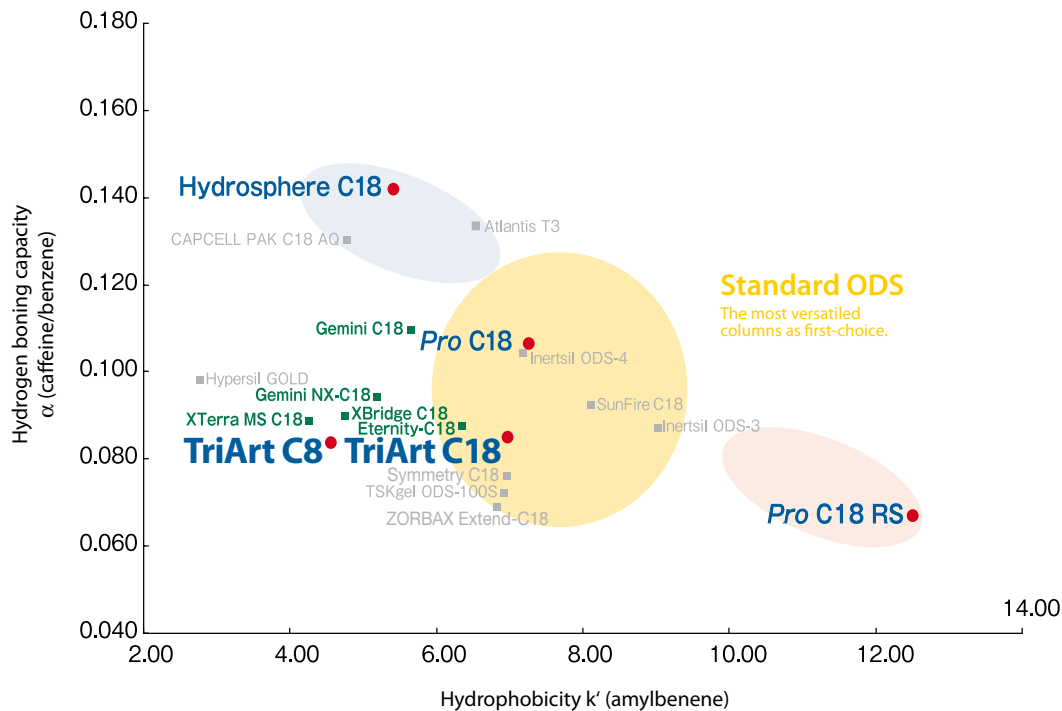


YMC-TriArt

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First choice column for method development



Conventional hybrid silica-based ODS columns tend to be less hydrophobic than silica-based columns. YMC-TriArt C18 has a higher carbon load, giving it a hydrophobicity comparable to that of standard ODS columns, thereby making it a "versatile first-choice" column for method development.

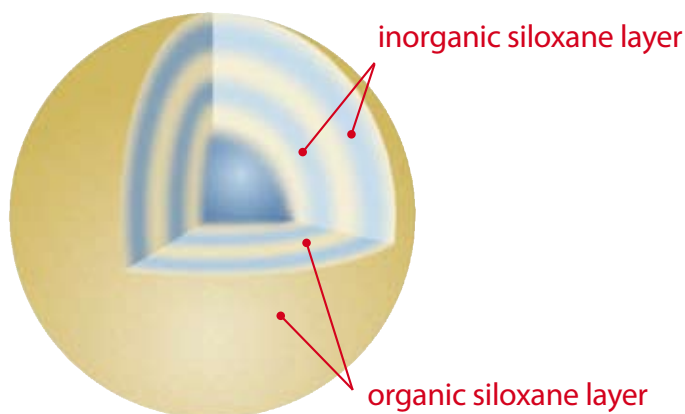
Chromatographers always seek to push the limits of HPLC columns to greater extremes to allow them to perform day-to-day with ever-changing pH, buffers and temperature ranges. The column for the laboratory of today must be suitable for harsh pH conditions in combination with high temperature ranges without sacrificing selectivity. In addition narrow, symmetrical peak shapes are necessary in order to cope with rapid analysis of demanding samples. This has required manufacturers to seek more innovative ways to produce suitable stationary phases.

In order to meet these goals, YMC has developed a new particle technology. This is based on a multi-layered particle produced via a tightly controlled granulation technology which has been adapted from micro-reactor technology. The revolutionary production technique provides a multi-layer silica-organic hybrid stationary phase, which provides an outstandingly narrow pore size and particle size distribution. This in turn, results in low back pressures and high loadability.

Particle technology

YMC-TriArt is a multi-layered material prepared using tightly controlled particle formation technology which has been adapted from micro-reactor technology. This recently developed production process results in exceptionally narrow particle and pore size distributions. With YMC-TriArt, challenging pH and high temperature conditions are no longer a limitation to the day-to-day work in laboratories. Most importantly, due to its unique particle composition, a balanced hydrophobicity and silanol activity are achieved which makes YMC-TriArt a "First Choice" column in method development.

YMC-TriArt hybrid structure

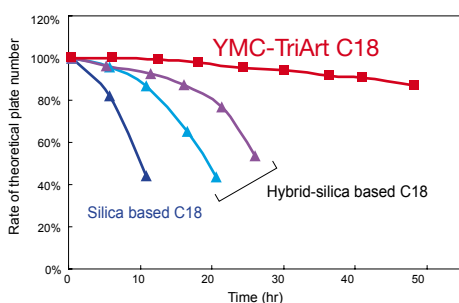


Specification

	YMC-TriArt C18	YMC-TriArt C8
Base	organic/inorganic silica	organic/inorganic silica
Stationary phase	C18 (as USP L1)	C8 (as USP L7)
Particle size	1.9, 3 and 5 μm	1.9, 3 and 5 μm
Pore size	12 nm	12 nm
Carbon load	16% (20% in total)	7% (11% in total)
Bonding	polymeric type	polymeric type
End-capping	multi-stage hybrid groups	multi-stage hybrid groups
pH range	1 ~ 12	1 ~ 12
Temperature range	pH 1-7: 70 °C, pH 7-12: 50 °C	pH 1-7: 70 °C, pH 7-12: 50 °C

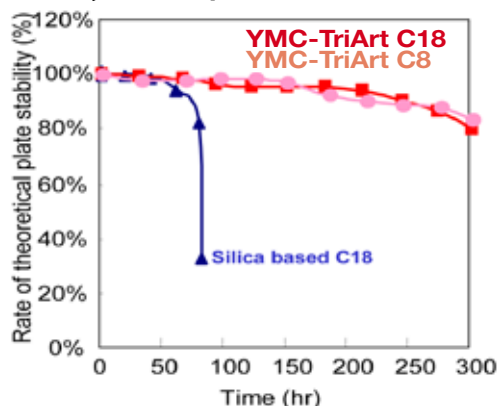
Versatile wide pH stability

Phosphate buffer (pH 11.5, 40 °C)



Column: 5 μ m, 150 x 4.6 mm ID
 Part-No.: TA12S051546PT
 Eluent: 50 mM K_2HPO_4 - K_2HPO_4 (pH 11.5) / methanol (90/10)
 Flow rate: 1.0 ml/min
 Temperature: 40 °C
 Sample: benzyl alcohol

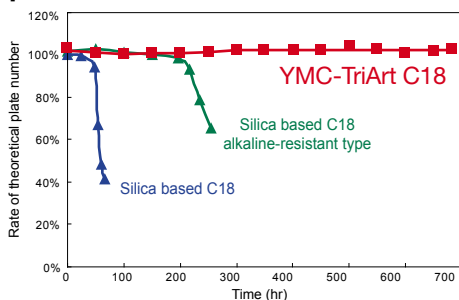
Triethylamine (pH 11.5, 40 °C)



Column: 5 μ m, 150 x 4.6 mm ID
 Part-No.: TA12S051546PT
 Eluent: 50 mM triethylamine (pH 11.5) / methanol (90/10)
 Flow rate: 1.0 ml/min
 Sample: benzyl alcohol

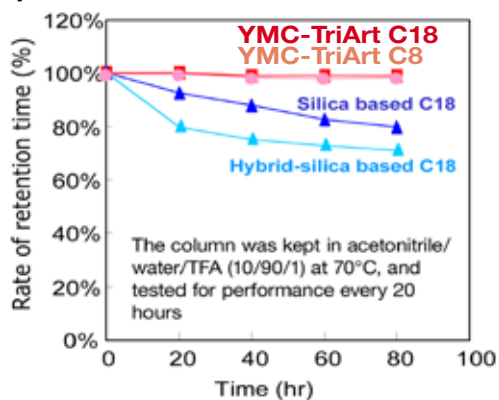
Durability at high temperature

pH 6.9, 70 °C



Column: 5 μ m, 50 x 2.0 mm ID
 Part-No.: TA12S050502PT
 Eluent: 20 mM KH_2PO_4 - K_2HPO_4 (pH 6.9) / acetonitrile (90/10)
 Flow rate: 0.2 ml/min
 Temperature: 70 °C
 Sample: phenol

pH 1, 70 °C



Column: 5 μ m, 50 x 2.0 mm ID
 Part-No.: TA12S050502PT
 Eluent: acetonitrile / water (60/40)
 Flow rate: 0.2 ml/min
 Temperature: 70 °C
 Sample: butyl benzoate

YMC-TriArt C18 and C8 show great chemical stability due to the newly developed hybrid-silica. Even under high pH or high temperature conditions, the lifetime of YMC-TriArt C18 and C8 is more than 10x greater than conventional reversed phase columns.

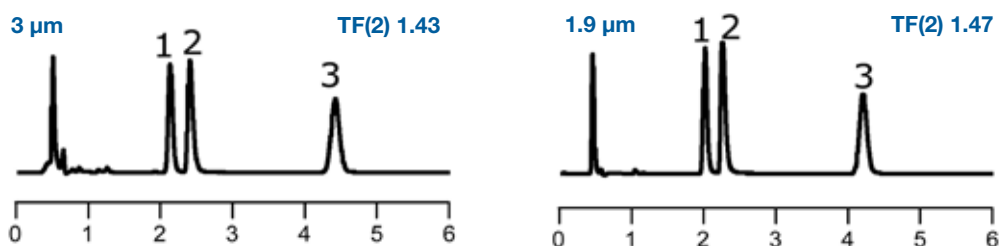
6

Transfer
HPLC
↕
UHPLC

Secure your method transfer!

Differences in selectivity, retention time, and also peak shapes between different particle sizes of commercially available C18 phases in the same brand (or an alternative as recommended by its manufacture) have been observed.

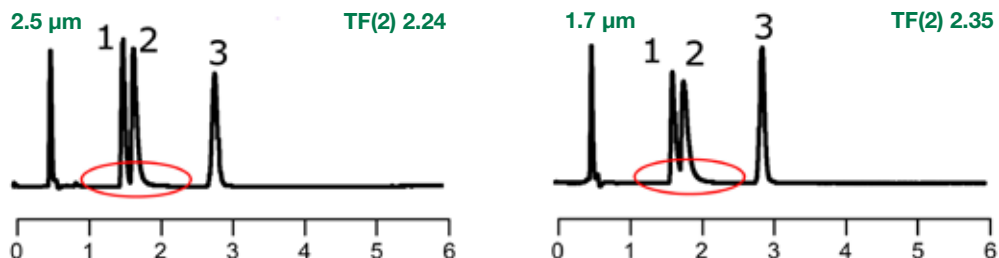
YMC-TriArt C18



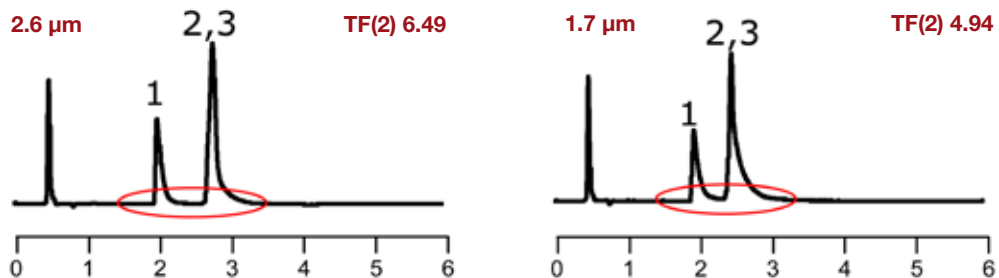
YMC has addressed this issue of method transfer. YMC-TriArt columns show identical selectivity and excellent peak shapes for basic compounds for all 3.0 µm to 1.9 µm particle sizes. It allows predictable scale up from UHPLC to conventional HPLC and even to semi-preparative LC, and vice versa.

Case Studies*

X-Bridge BEH C18 and Acquity UPLC BEH C18



Kinetex™ C18

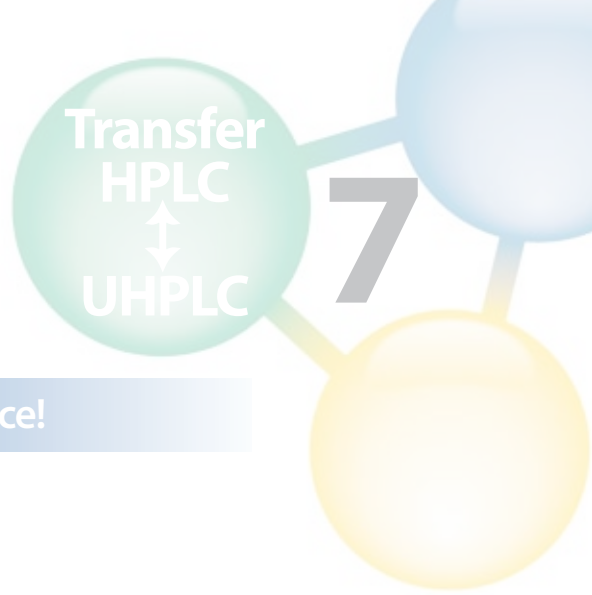


Kinetex™ C18 columns show significant peak tailing™ and have limited scalability due to lack of larger particle sizes.

Column: 50 x 2.0 mm ID or 2.1 mm ID
Eluent: 20 mM KH_2PO_4 - K_2HPO_4 (pH 6.9) / acetonitrile (65/35)
Temperature: 40 °C
Flow rate: 0.2 ml/min
Detection: UV at 235 nm

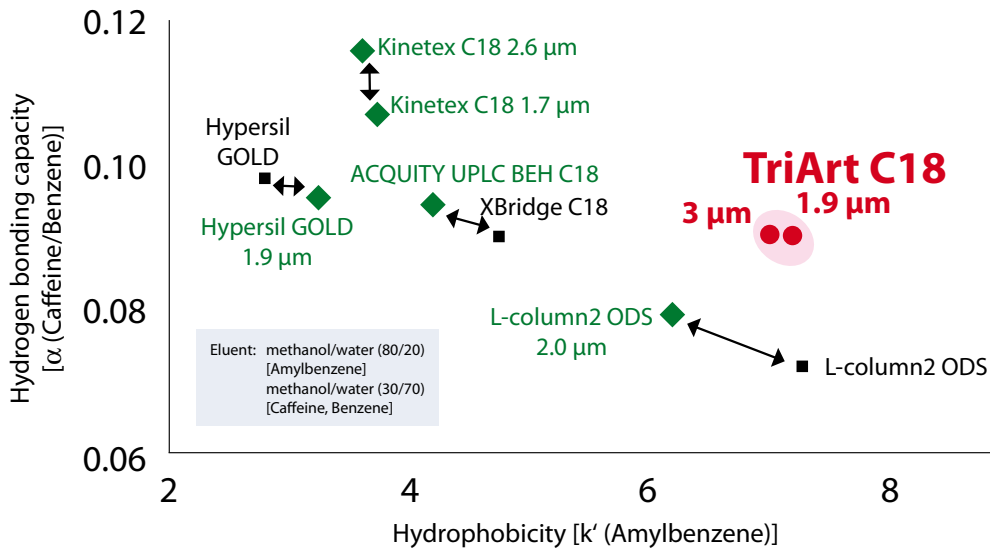
1. Chlorpheniramine (basic)
2. Dextromethorphan (basic)
3. Propyl paraben (internal standard)

* These observations might not be representative for all applications but have been reported in some cases.



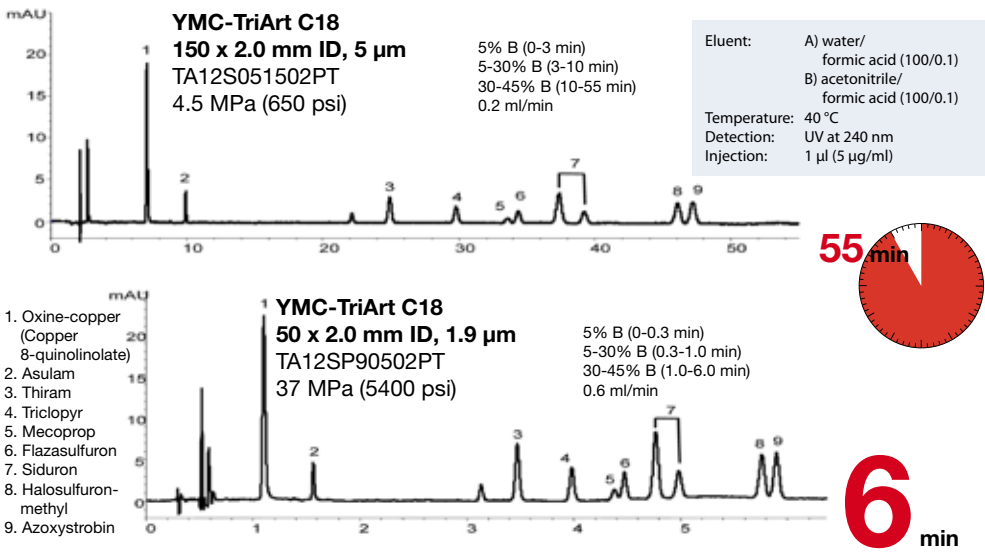
Transfer
HPLC
↕
UHPLC

Evaluation of method transfer performance!



With the introduction of UHPLC, sub-2- μ m particles became necessary. Therefore smaller particles have been added to existing column lines. Consequently, sub-2- μ m particles may exhibit differences in chromatographic performance. By introducing YMC-TriArt, YMC provides matching chromatographic behaviour for **all** particles sizes!

Method transfer between HPLC ↔ UHPLC



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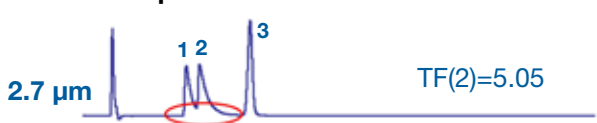
UHPLC

Higher resolution and good loadability

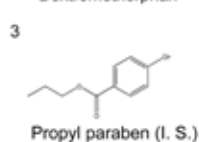
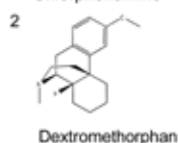
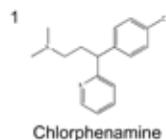
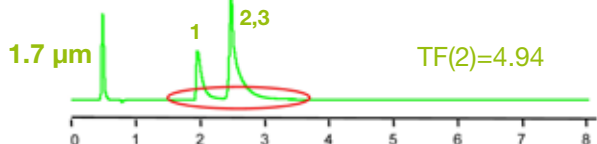
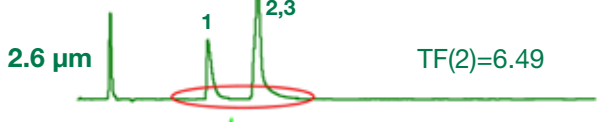
YMC-TriArt C18



Ascentis Express C18

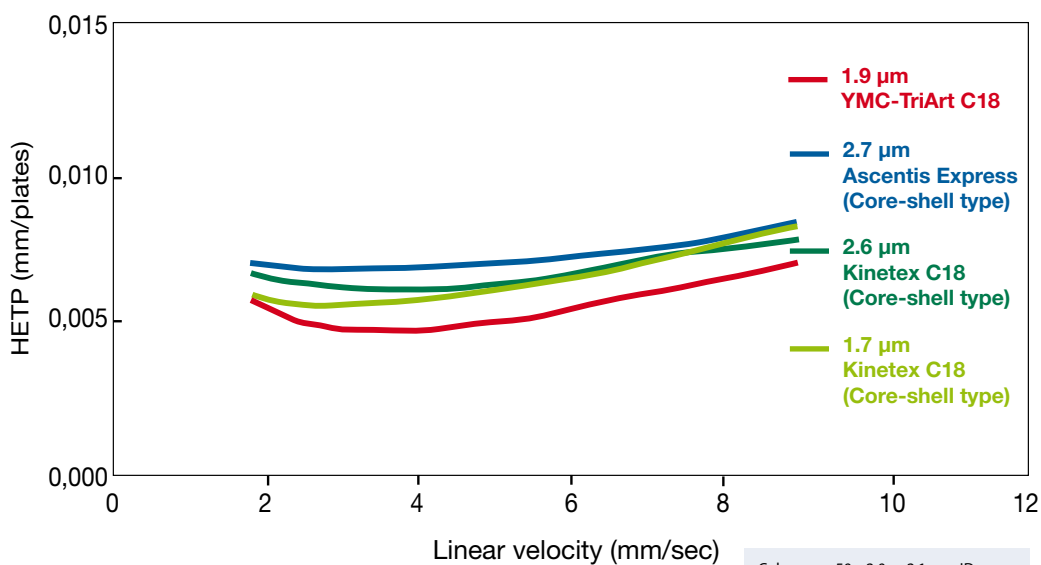


Kinetex C18



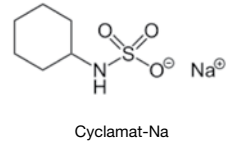
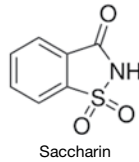
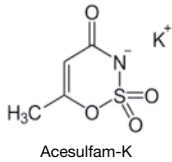
Column: 50 x 2.0 or 2.1 mm ID
 Eluent: 20 mM KH₂PO₄-K₂HPO₄ (pH 6.9)/ acetonitrile (65/35)
 Flow rate: 0.2 ml/min
 Detection: UV at 235 nm
 Temperature: 40 °C

Lower HETP means higher resolution!

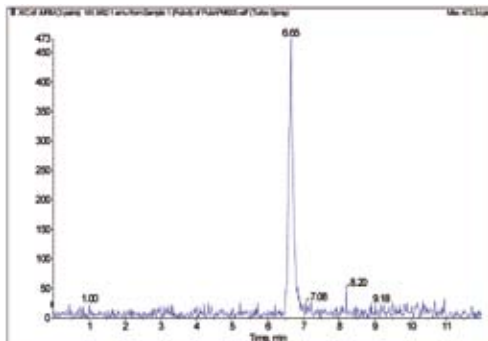


Column: 50 x 2.0 or 2.1 mm ID
 Eluent: acetonitrile / water (60/40)
 Detection: UV at 254 nm
 Sample: Butylbenzoate

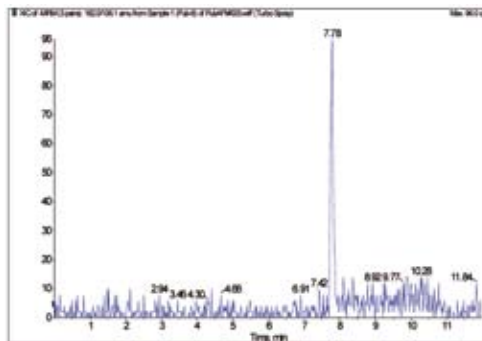
Determination of Artificial Sweeteners with LC-MS/MS



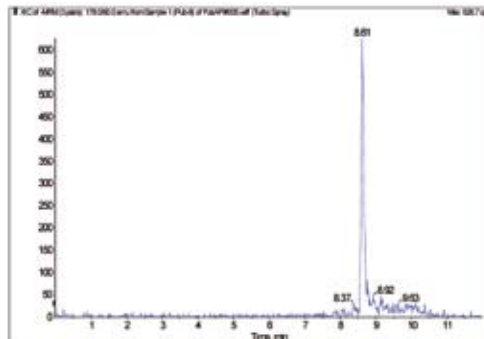
→ Non biological markers of wastewater entries in ground and surface water



Extracted Ion Chromatogram (XIC) of Acesulfam-K, 0.1 µg/L



Extracted Ion Chromatogram (XIC) of Saccharin, 0.1 µg/L



Extracted Ion Chromatogram (XIC) of Cyclamat-Na, 0.1 µg/L

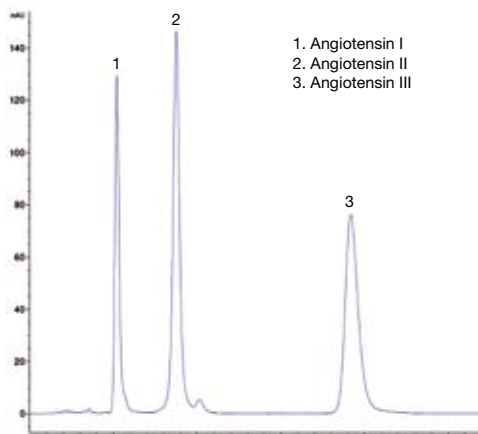
Column: YMC-TriArt C18, 12 nm, 1.9 µm, 100 x 3.0 mm ID
 Part-No.: TA12SP91003PT
 LC-System: Agilent 1100 HPLC system and CTC Analytics
 HTC-Pal Autosampler
 MS/MS System: Applied Biosystems MDS Sciex API 4000,
 ESI negative
 Temperature: 35°C
 Flow: 0.3 ml/min
 Injection: 40 µL, direct injection
 Eluent: A: H₂O (containing 10 mmol NH₄ formate)
 B: MeOH (containing 10 mmol NH₄ formate)
 Gradient:

Time	0	6.0	6.1	12.0
% B	2	75	2	2

by courtesy of: Thomas Class, Sandro Jooß
 PTRL Europe, Helmholtzstraße 22, Science Park I, D-89081 Ulm

10 UHPLC

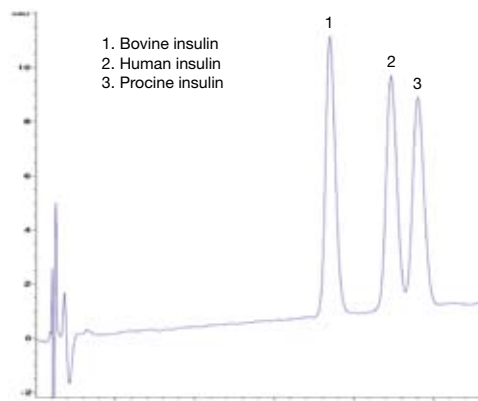
Angiotensin I, II and III



1. Angiotensin I
2. Angiotensin II
3. Angiotensin III

Column: TA12SP90502PT
 Dimension: 50 x 2.0 mm ID
 Eluent: 20 mM KH_2PO_4 + K_2HPO_4 (pH 7.9) / acetonitrile (22/78)
 Flow rate: 0.7 ml/min
 Detection: UV at 220 nm
 Pressure: 720 bar
 Injection: 0.5 μl
 Temperature: 40 °C

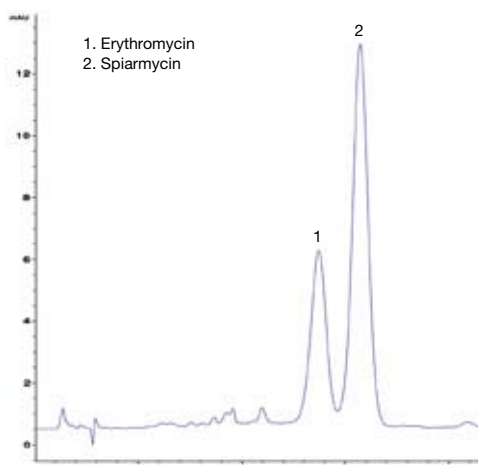
Insulin



1. Bovine insulin
2. Human insulin
3. Procine insulin

Column: TA12SP90502PT
 Dimension: 50 x 2.0 mm ID
 Eluent: A) H_2O + 0.1% TFA
 B) acetonitrile + 0.1% TFA
 Gradient: 30% B (0 min); 30-32% B (0-5 min); 32% B (55 min)
 Flow rate: 0.6 ml/min
 Detection: UV at 220 nm
 Pressure: 611 bar
 Injection: 0.5 μl
 Temperature: 30 °C

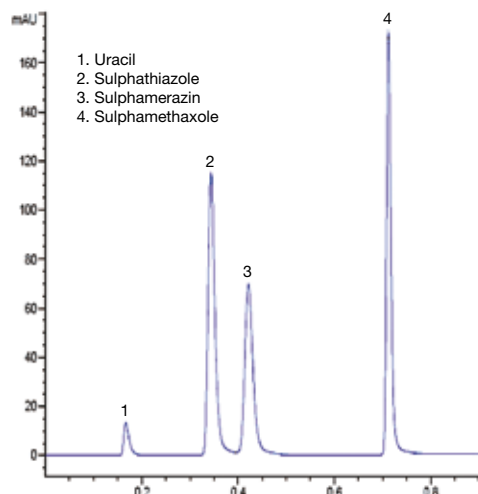
Macrolide antibiotics



1. Erythromycin
2. Spiarmycin

Column: TA12SP90502WT
 Dimension: 50 x 2.0 mm ID
 Eluent: A) 20 mM K_2HPO_4 + 20 mM KH_2PO_4 (pH 7.9)
 B) acetonitrile
 Gradient: 60% B (0.5 min); 60-70% B (0.5-1.5 min); 70% B (3.5 min)
 Flow rate: 0.45 ml/min
 Detection: UV at 210 nm
 Pressure: 520 bar
 Injection: 1 μl
 Temperature: 50 °C

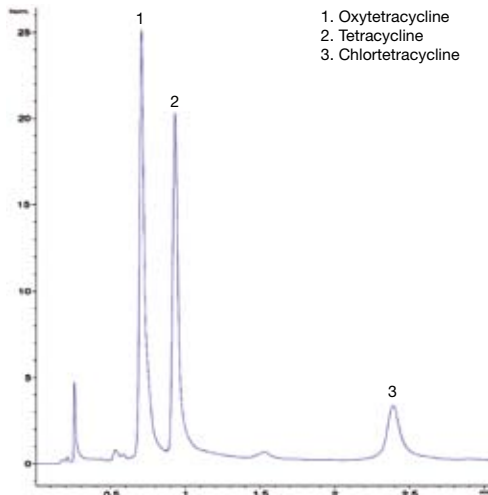
Sulpha drugs



1. Uracil
2. Sulphathiazole
3. Sulphamerazin
4. Sulphamethaxole

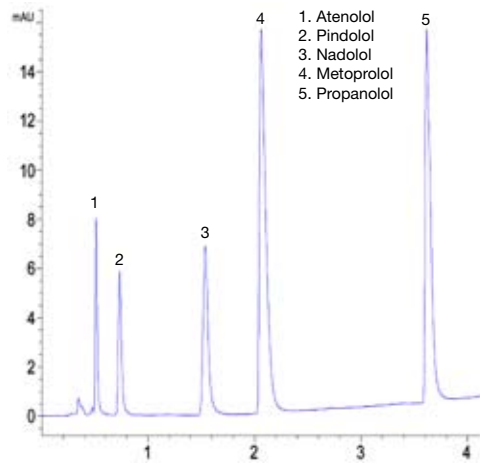
Column: TA12SP90502WT
 Dimension: 50 x 2.0 mm ID
 Eluent: H_2O + formic acid (pH 2.5) / acetonitrile (75/25)
 Flow rate: 0.75 ml/min
 Detection: UV at 280 nm
 Pressure: 740 bar
 Injection: 0.5 μl
 Temperature: 50 °C

Tetracycline antibiotics



Column: TA12SP90502WT
Dimension: 50 x 2.0 mm ID
Eluent: 5 mM $\text{NH}_4\text{CH}_2\text{COOH}$ / acetonitrile (87/13)
Flow rate: 0.65 ml/min
Detection: UV at 280 nm
Pressure: 662 bar
Injection: 1 μl
Temperature: 40 °C

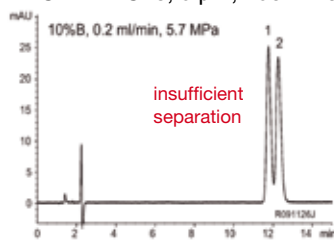
Betablockers



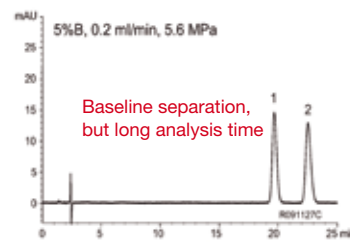
Column: TA12SP90502PT
Dimension: 50 x 2.0 mm ID
Eluent: A) 20 mM $\text{NH}_4\text{CH}_2\text{COOH}$ + ammonia (pH 9.0)
B) acetonitrile
Gradient: 25% B (1.0 min); 75% B (1-6 min)
Flow rate: 0.35 ml/min
Detection: UV at 254 nm
Pressure: 450 bar
Injection: 1 μl
Temperature: 40 °C

Fast LC for conventional HPLC

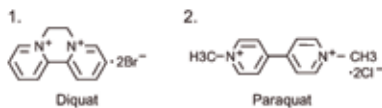
YMC-TriArt C18, 5 μm , 150 x 2.0 mm ID



optimisation

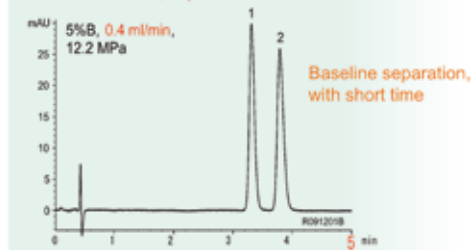


Down scaling



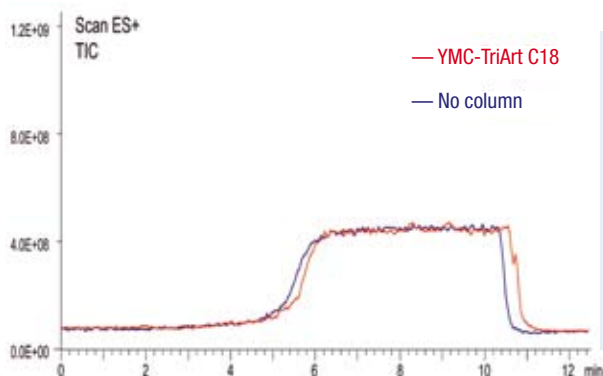
Eluent: A) water / HFBA* (100/0.1)
B) acetonitrile / HFBA* (100/0.1)
Temperature: 37 °C
Detection: UV at 290 nm
Injection: 1 μl (0.1 mg/ml)
*heptafluorobutyric acid

YMC-TriArt C18, 3 μm , 50 x 2.0 mm ID



12 LC/MS

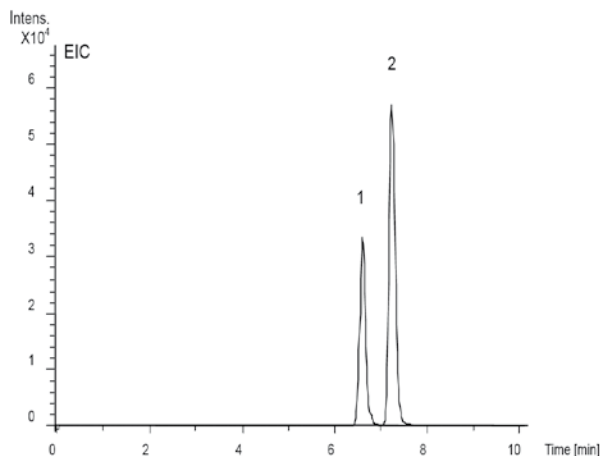
LC/MS compatibility



Column: 5 μ m, 50 x 2.0 mm ID
 Part-No.: TA12S050502PT
 Eluent: A) water / formic acid (100/0.1)
 B) acetonitrile / formic acid (100/0.1)
 5% B (0-1 min), 5-100% B (1-5 min), 100% B (5-10 min), 100-5% B (10-10.1 min), 5% B (10.1-12.5 min)
 Flow rate: 0.4 ml/min
 Temperature: 40 °C
 Detection: ESI positive, TIC (Mass Range: 50-1000)

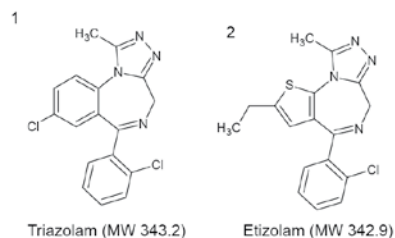
Column bleeding, caused by the fragments of stationary phase, is the main reason for background noise and restrictions on detection limits. No bleed is observed in the test of total ion current (TIC) measured by LC/MS with blank or with YMC-TriArt C18. So in terms of the signal/noise ratio (S/N ratio), YMC-TriArt C18 can be expected to not only reduce the background noise but to also increase the sensitivity of the analysis.

LC/MS analysis of benzodiazepine derivatives



Peak 1: 100ng_mL_RB4_01_2291.d: EIC 343.051±0.01+All MS
 Peak 2: 100ng_mL_RB4_01_2291.d: EIC 343.078±0.01+All MS

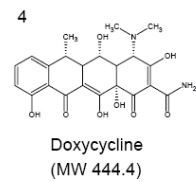
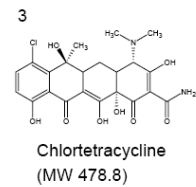
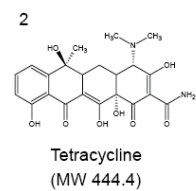
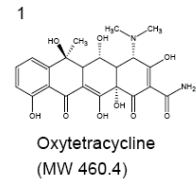
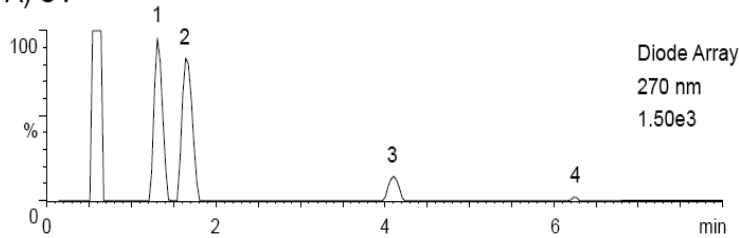
Courtesy of J. Watanabe, Bruker Daltonics K. K.



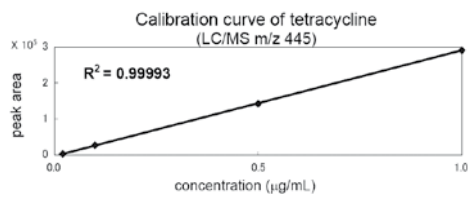
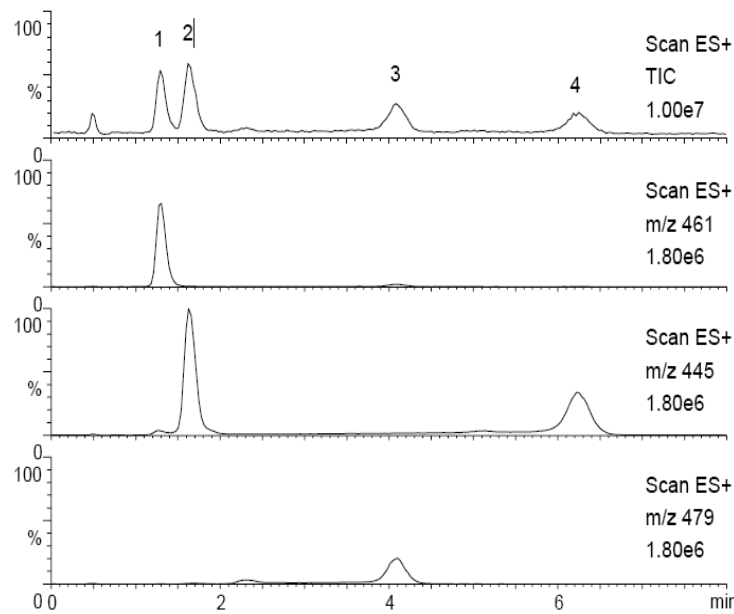
Column: YMC-TriArt C18 (5 μ m, 12 nm)
 50 x 2.0 mm ID
 Part-No.: TA12S050502PT
 Eluent: A) 10 mM formic acid
 B) acetonitrile
 Gradient: 25-50% B (0-10 min)
 Flow rate: 0.2 ml/min
 Temperature: 40 °C
 Detection: Bruker Daltonics micrOTOF, ESI, positive mode
 Injection: 5 μ l (100 ng/ml)

LC/MS analysis of tetracycline antibiotics

A) UV



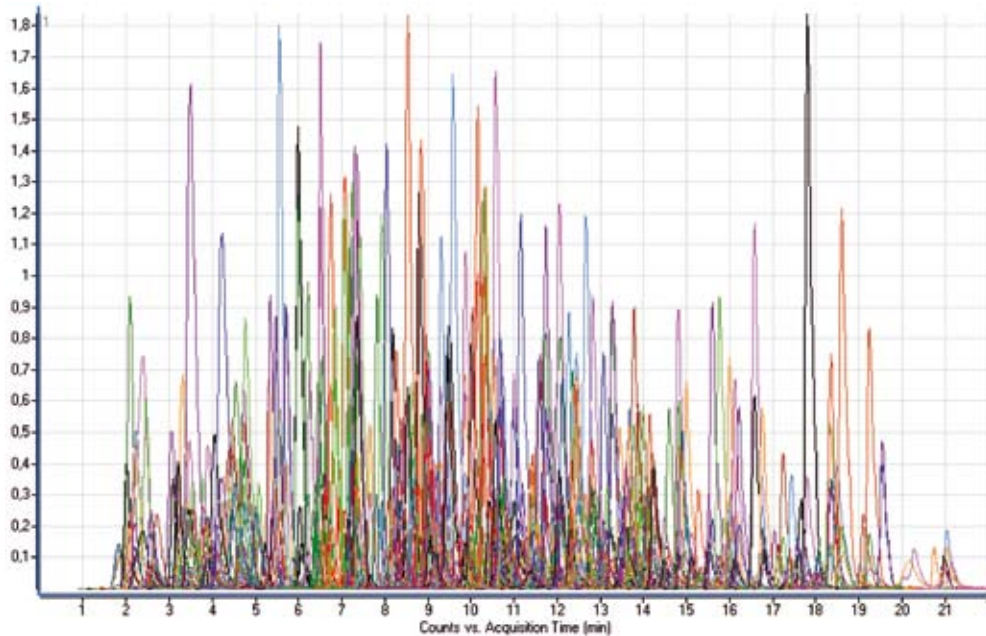
B) MS



Column: YMC-TriArt C18 (5 μm , 12 nm) 50 x 2.0 mm ID
 Part-No.: TA12S050502PT
 Eluent: acetonitrile / water / formic acid (15/85/0.1)
 Flow rate: 0.4 ml/min
 Temperature: 40 °C
 Detection: A) UV at 270 nm
 B) ESI positive-mode
 Injection: 10 μl (1 $\mu\text{g/ml}$)

14 LC/MS

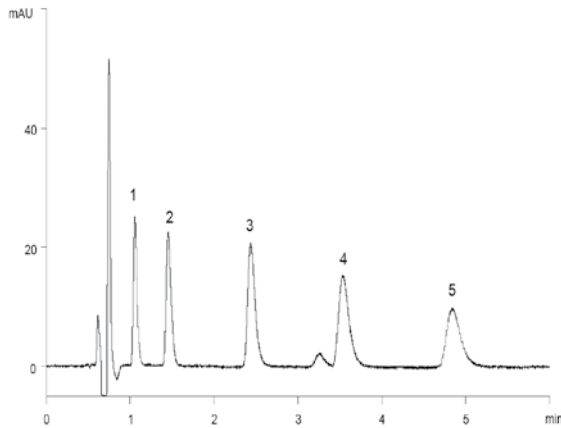
Analysis of 360 pesticides in a single run



Column:	YMC-TriArt C18 (3 μ m, 100 x 2.0 mm ID)	Injection:	5 μ l
Part-No.:	TA12S031002PT	Gradient:	0 min: 30% B, 0.1 min: 50% B, 18 min: 100% B, 21 min: 100% B, 21.01 min: 30% B, 29 min: 30% B
Eluent:	A) 5 mM ammonium formate / water B) 5 mM ammonium formate / methanol	Total run time:	30 min
Flow rate:	0.25 ml/min	Detection:	
Temperature:	45 $^{\circ}$ C	Sample:	100 ng/ml pesticide mix in acetonitrile

by courtesy of: József László
WIREC, WESSLING International Research and Educational Centre Nonprofit Co. (Hungary)

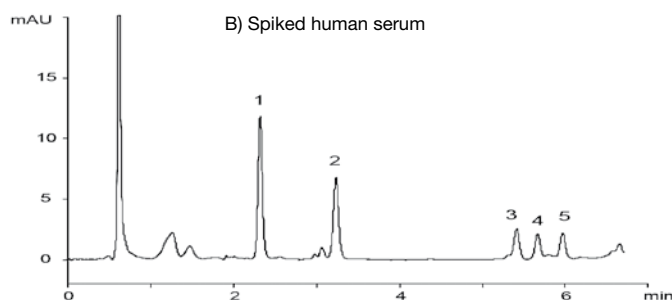
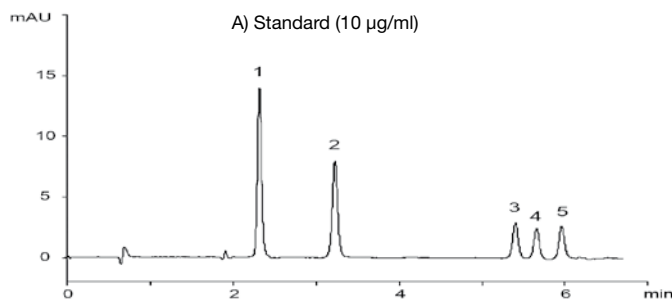
Separation of alkaloids



1. Scopolamine
2. Atropine
3. Cinchonine
4. Quinine
5. Dihydroquinine

Column: YMC-TriArt C18 (5 μ m, 12 nm)
50 x 2.0 mm ID
Part-No.: TA12S050502PT
Eluent: 20 mM CH₃COOH-CH₃COONH₄
(pH 4.9) / acetonitrile (80/20)
Flow rate: 0.2 ml/min
Temperature: 40 °C
Detection: UV at 220 nm
Injection: 1 μ l (0.02-0.1 mg/ml)

Barbiturates in human serum



Solid-phase extraction method

YMC Dispo SPE C18 100 mg/1ml

Condition

2 ml methanol
2 ml water

Load

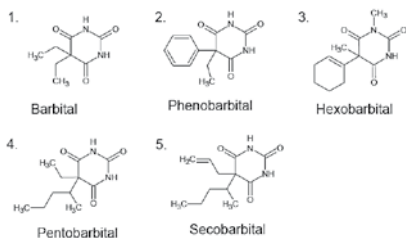
500 μ l spiked human serum
solution (each 10 μ g)

Elute

500 μ l methanol/water (85/15)

Dilute

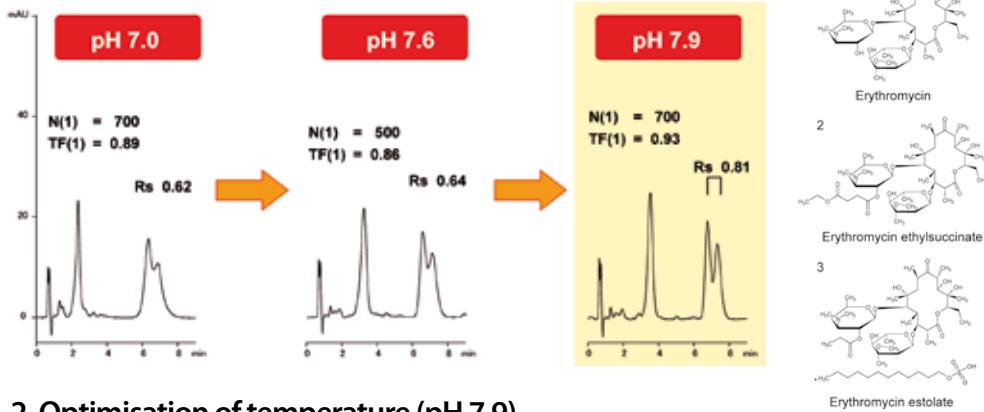
500 μ l 20 mM ammonium
formate buffer (pH 9.5)



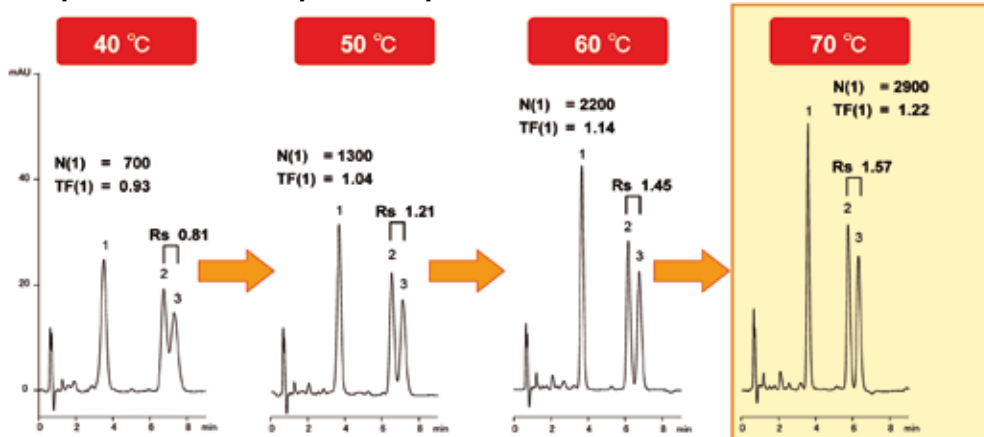
Column: YMC-TriArt C18 (5 μ m, 12 nm)
50 x 2.0 mm ID
Part-No.: TA12S050502PT
Eluent: A) 20 mM HCOONH₄-NH₃ (pH 9.5)
B) methanol
Gradient: 0-90% B (0-7 min)
Flow rate: 0.2 ml/min
Temperature: 25 °C
Detection: UV at 240 nm
Injection: 1 μ l

Erythromycin at elevated pH and temperature

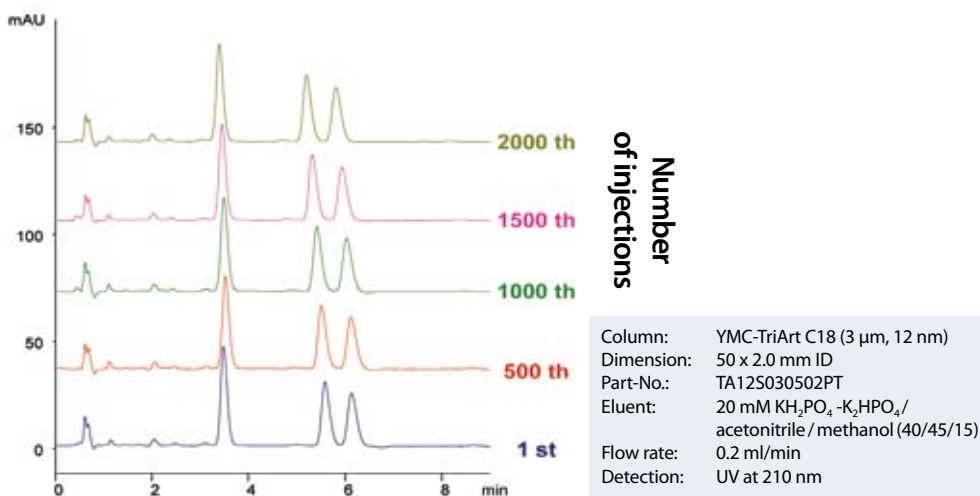
1. Optimisation of pH



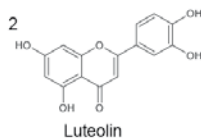
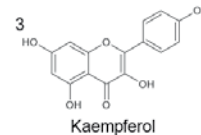
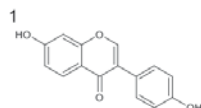
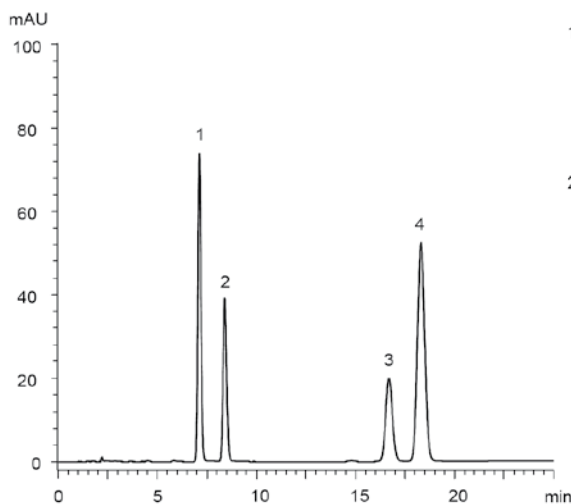
2. Optimisation of temperature (pH 7.9)



3. Stability test: pH 7.9, 70 °C

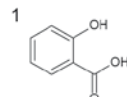
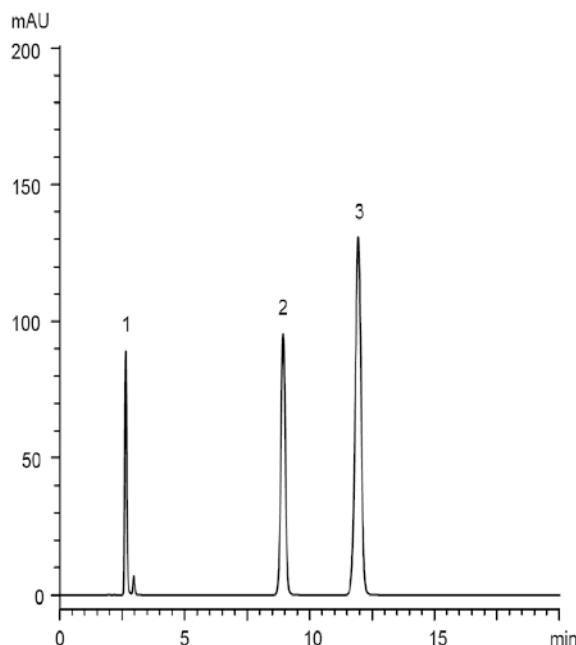


Separation of flavonoids

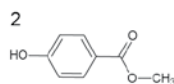
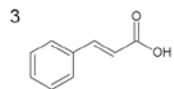


Column: YMC-TriArt C18 (5 μ m, 12 nm)
Dimension: 150 x 3.0 mm ID
Eluent: acetonitrile / 10 mM H_3PO_4 (30/70)
Flow rate: 0.425 ml/min
Temperature: 37 $^\circ\text{C}$
Detection: UV at 280 nm
Injection: 2 μ l (50 $\mu\text{g}/\text{ml}$)

Separation of aromatic carboxylic acids

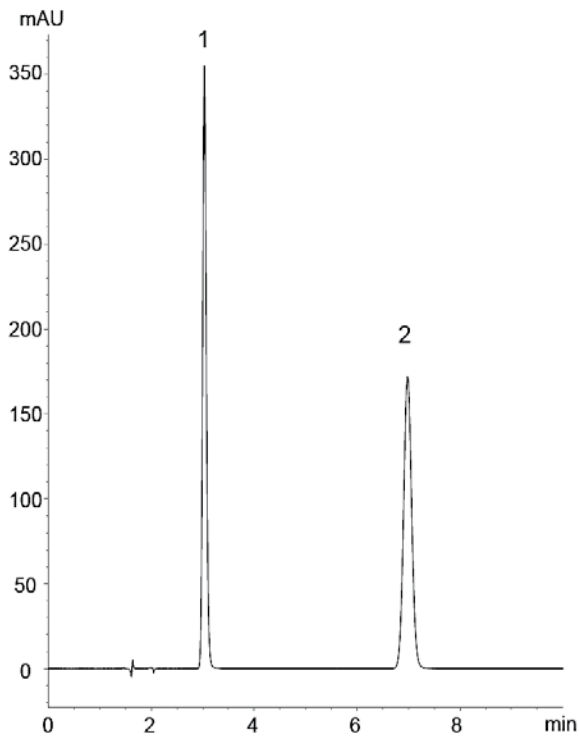


Salicylic acid

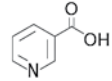
Methyl *p*-hydroxybenzoate (I.S.)*trans*-Cinnamic acid

Column: YMC-TriArt C18 (5 μ m, 12 nm)
Dimension: 150 x 3.0 mm ID
Part-No.: TA12S051503PT
Eluent: 10 mM $\text{CH}_3\text{COOH}-\text{CH}_3\text{COONH}_4$ (pH 4.2) / acetonitrile (75/25)
Flow rate: 0.425 ml/min
Temperature: 40 $^\circ\text{C}$
Detection: UV at 254 nm
Injection: 4 μ l (0.02 ~ 0.3 mg/ml)

Aciclovir syrup and injection

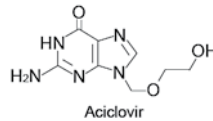


1.



Nicotinic acid

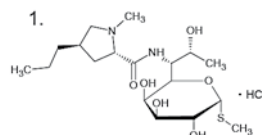
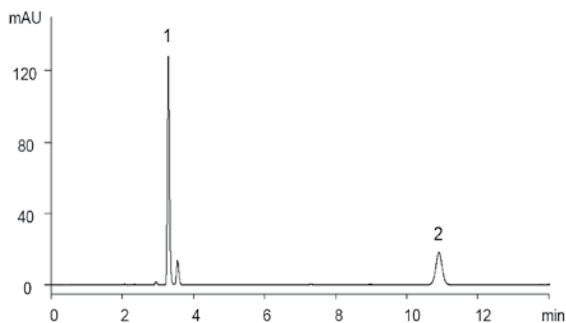
2.



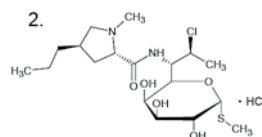
Aciclovir

Column: YMC-TriArt C18 (5 μ m, 12 nm)
150 x 4.6 mm ID
Part-No.: TA12S051546PT
Eluent: phosphate buffer* / methanol (95/5)
*Dissolve 1.45 g of H₃PO₄ and 25 ml of 1 mol/l CH₃COOH in water to make 900 ml → adjust pH 2.5 by 1 mol/l NaOH → add water to make 1000 ml
Flow rate: 1.0 ml/min
Temperature: 25 °C
Detection: UV at 254 nm
Injection: 20 μ l (0.05 mg/ml, 0.032 mg/ml)

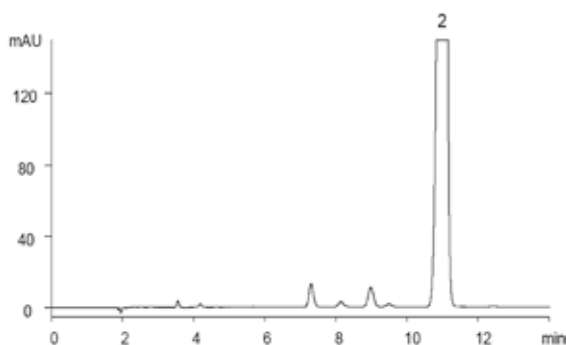
Clindamycin hydrochloride



Lincomycin hydrochloride

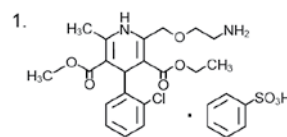
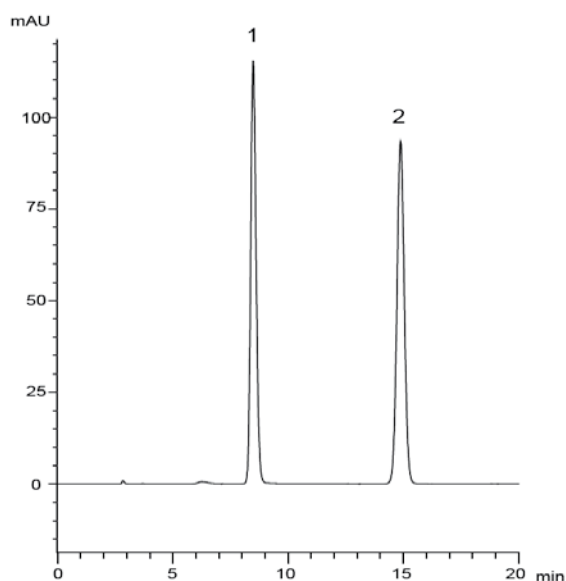


Clindamycin hydrochloride

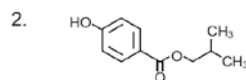


Column: YMC-TriArt C18 (5 μ m, 12 nm)
 Dimension: 250 x 4.6 mm ID
 Part-No.: TA12S052546PT
 Eluent: 50 mM KH_2PO_4 (pH 7.5 adjusted by 8 M KOH) / acetonitrile (55/45)
 Flow rate: 1.0 ml/min
 Temperature: 25 $^\circ\text{C}$
 Detection: UV at 210 nm
 Injection: 10 μ l

Analysis of amlodipine besilate



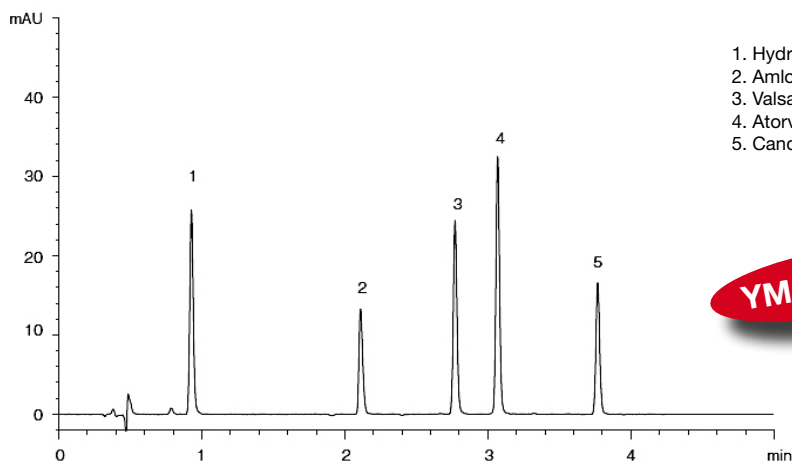
Amlodipine besilate



Isobutyl *p*-hydroxybenzoate

Column: YMC-TriArt C18 (5 μ m, 12 nm)
 Dimension: 150 x 3.0 mm ID
 Part-No.: TA12S051503PT
 Eluent: 10 mM $\text{CH}_3\text{COOH}-\text{CH}_3\text{COONH}_4$ (pH 4.2) / acetonitrile (75/25)
 Flow rate: 0.425 ml/min
 Temperature: 40 $^\circ\text{C}$
 Detection: UV at 254 nm
 Injection: 4 μ l (0.02 ~ 0.3 mg/ml)

Basic drugs

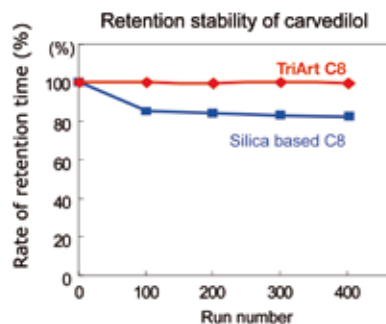
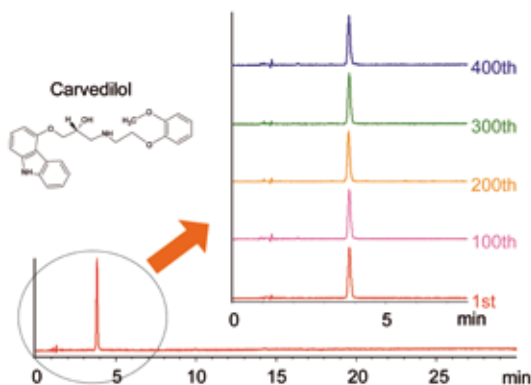


1. Hydrochlorothiazide
2. Amlodipine besilate
3. Valsartan
4. Atorvastatin calcium hydrate
5. Candesartan cilexetil

YMC-TriArt C8

Column:	YMC-TriArt C8 (3 μ m, 12 nm), 50 x 2.0 mm ID	Flow rate:	0.4 ml/min
Part-No.:	TO12S030502PT	Temperature:	30 $^{\circ}$ C
Eluent:	A) water / formic acid (100/0.1) B) acetonitrile / formic acid (100/0.1) 10-90% B (0-5 min), 90% B (5-7 min)	Detection:	UV at 254 nm
		Injection:	2 μ l (10-20 μ g/ml)

Sequential analysis of Carvedilol

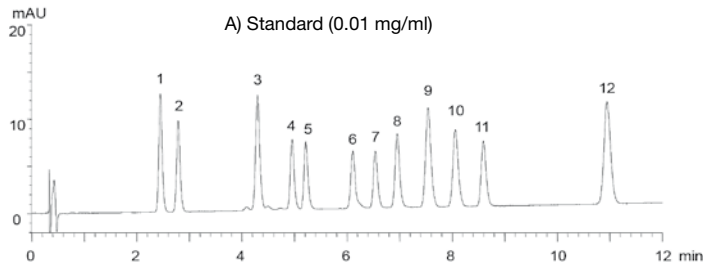


YMC-TriArt C8

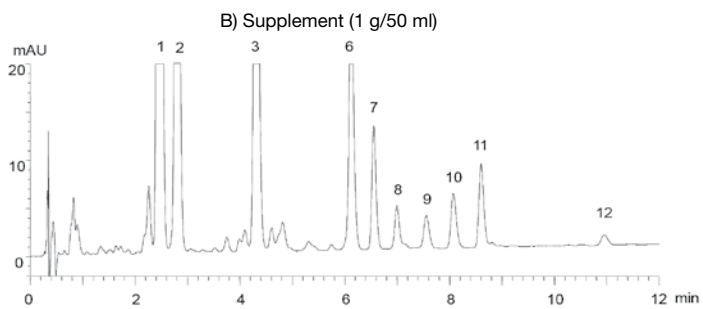
Column:	YMC-TriArt C8 (5 μ m, 150 x 2.0 mm ID)
Part-No.:	TO12S051502PT
Eluent:	phosphate buffer (pH 2.0)* / acetonitrile (65/35) *Dissolve 2.72 g of KH_2PO_4 in 900 ml water, adjust pH 2.0 with H_3PO_4 , and add water to make 1000 ml
Flow rate:	0.28 ml/min (adjust the flow rate so that the retention time of carvedilol is about 4 min)
Temperature:	55 $^{\circ}$ C
Detection:	UV at 240 nm

No change in retention time is observed even under a high pH and at a elevated temperature.

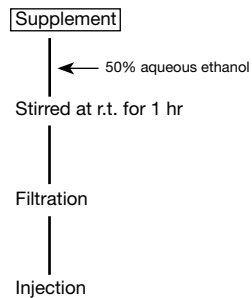
Soy isoflavones in supplement



1. Daidzin
2. Glycitin
3. Genistin
4. 6^{''}-O-Malonyldaidzin
5. 6^{''}-O-Malonylglycitin
6. 6^{''}-O-Acetyldaidzin
7. 6^{''}-O-Acetylglycitin
8. 6^{''}-O-Malonylgenistin
9. Daidzein
10. Glycitein
11. 6^{''}-O-Acetylgenistin
12. Genistein

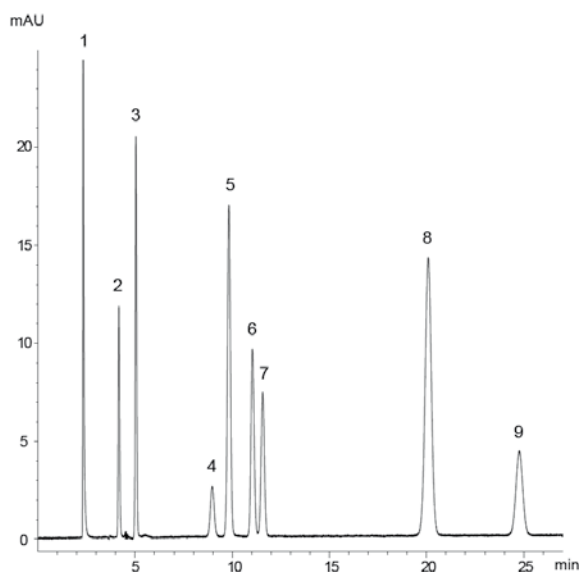


Sample preparation method



Column: YMC-TriArt C18 (3 μm, 12 nm) 50 x 2.0 mm ID Flow rate: 0.4 ml/min
 Eluent: A) acetonitrile / water / HCOOH (10/90/0.1) Temperature: 25 °C
 B) acetonitrile / water / HCOOH (60/40/0.1) Detection: UV at 254 nm
 Gradient: 5-40% B (0-12 min) Injection: 2 μl

Separation of water-soluble vitamins

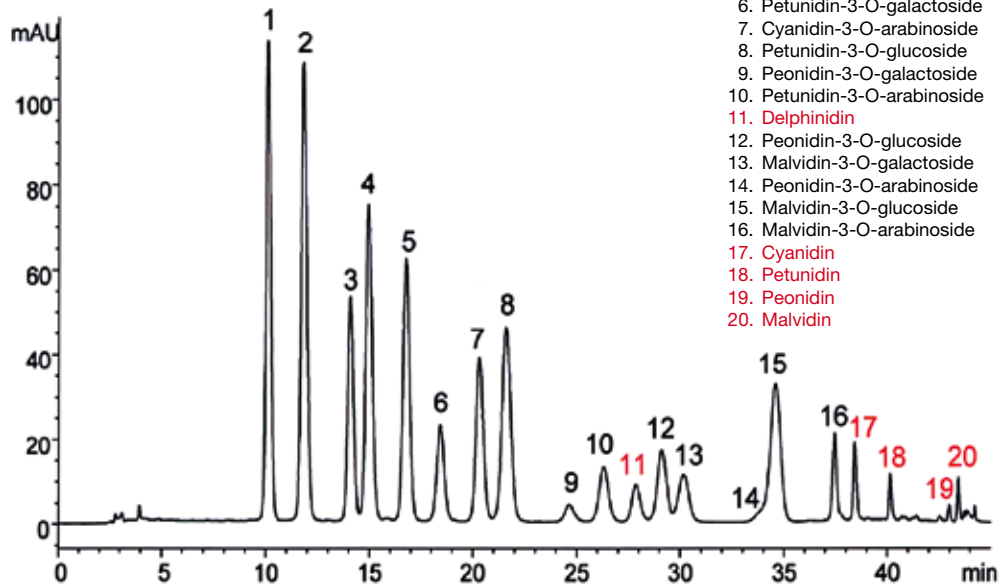


1. Thiamine HCl (Vitamin B₁)
2. Pyridoxine HCl (Vitamin B₆)
3. Nicotinamide
4. Cyanocobalamin (Vitamin B₁₂)
5. L-Ascorbic acid 2-glucoside
6. L-Ascorbic acid (Vitamin C)
7. Erythorbic acid
8. Riboflavin (Vitamin B₂)
9. Nicotinic acid

Column: YMC-TriArt C18 (5 μm, 12 nm)
 250 x 4.6 mm ID
 Part-No.: TA12S052546PT
 Eluent: phosphate buffer* / acetonitrile (90/10)
 *Dissolve 1.4g KH₂PO₄ in 800ml water → add 26 ml 10% TBA-OH → adjust pH 5.2 by 20% H₃PO₄ → add water to make 1000 ml
 Flow rate: 0.8 ml/min
 Temperature: 40 °C
 Detection: UV at 260 nm
 Injection: 10 μl (5 μg/ml)

Analysis of anthocyanins and anthocyanidins

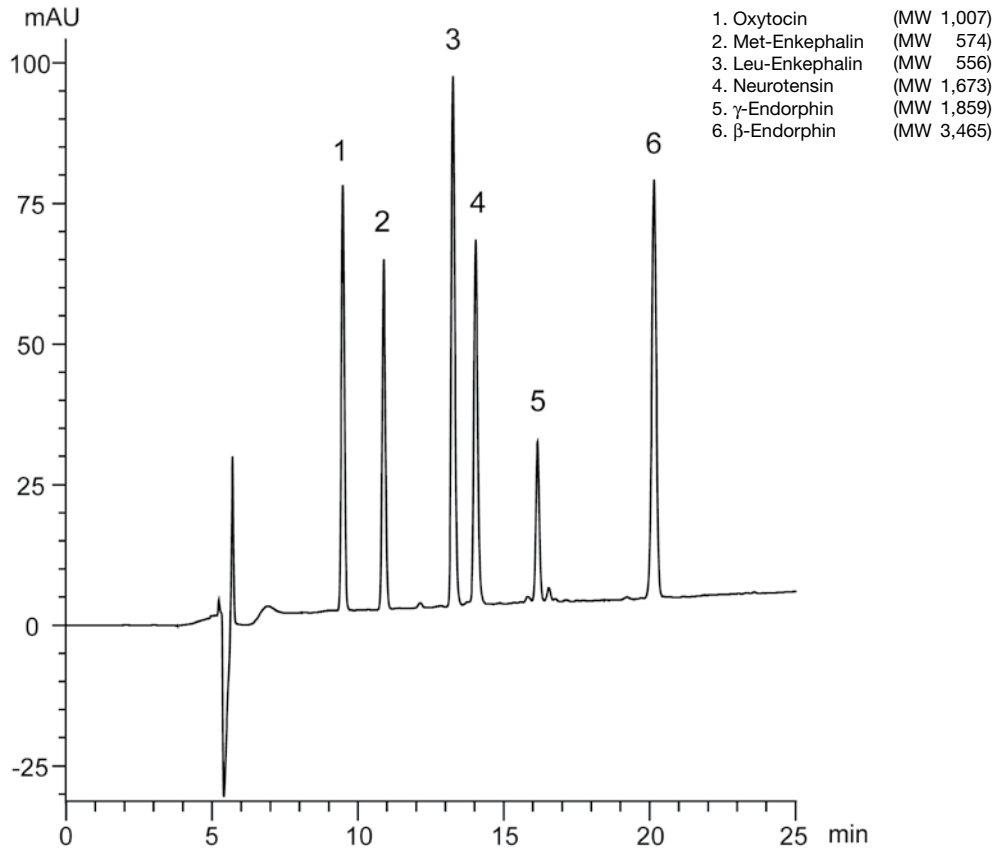
Anthocyanins: Indicated in black
 Anthocyanidins: Indicated in red



1. Delphinidin-3-O-galactoside
2. Delphinidin-3-O-glucoside
3. Cyanidin-3-O-galactoside
4. Delphinidin-3-O-arabinoside
5. Cyanidin-3-O-glucoside
6. Petunidin-3-O-galactoside
7. Cyanidin-3-O-arabinoside
8. Petunidin-3-O-glucoside
9. Peonidin-3-O-galactoside
10. Petunidin-3-O-arabinoside
11. Delphinidin
12. Peonidin-3-O-glucoside
13. Malvidin-3-O-galactoside
14. Peonidin-3-O-arabinoside
15. Malvidin-3-O-glucoside
16. Malvidin-3-O-arabinoside
17. Cyanidin
18. Petunidin
19. Peonidin
20. Malvidin

Column: YMC-TriArt C18 (5 μ m, 12 nm) 250 x 4.6 mm ID
 Part-No.: TA12S052546PT
 Eluent: A) water / formic acid (90/10)
 B) acetonitrile / methanol / water / formic acid (22.5/22.5/40/10)
 Gradient: 20-28% B (0-30 min), 28-70% B (30-40 min), 100% B (40-45 min)
 Flow rate: 1.0 ml/min
 Temperature: 25 $^{\circ}$ C
 Detection: UV/VIS at 535 nm
 Sample: commercial bilberry powder (1.25 mg/ml)

Peptides (MW 556 - 3,465)

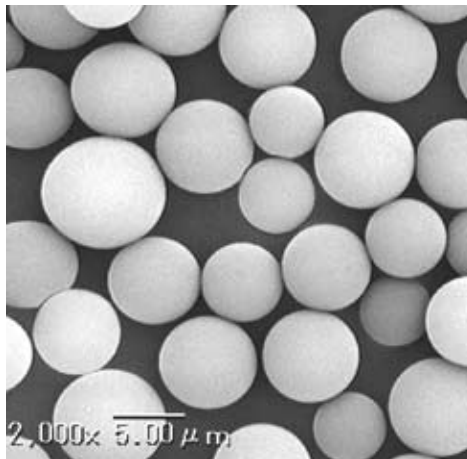


Column:	YMC-TriArt C18 (5 μm, 12 nm)	Flow rate:	0.2 ml/min
Dimension:	150 x 2.0 mm ID	Temperature:	37 °C
Part-No.:	TA12S051502PT	Detection:	UV at 220 nm
Eluent:	A) water / TFA (100/0.1) B) acetonitrile / TFA (100/0.1) 20-45% B (0-25 min)	Injection:	2 ml (0.075 - 0.25 mg/ml)

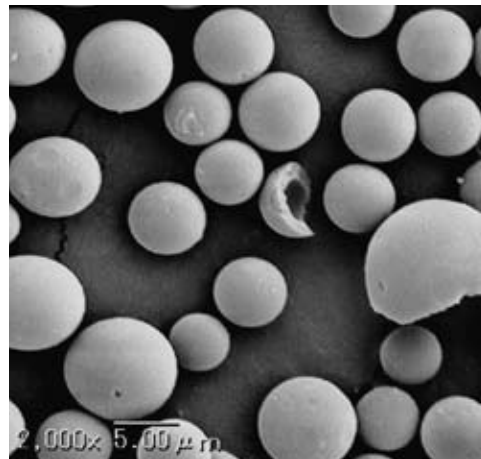
YMC-TriArt: Improved quality of particles

Uniform spherical particles

YMC-TriArt

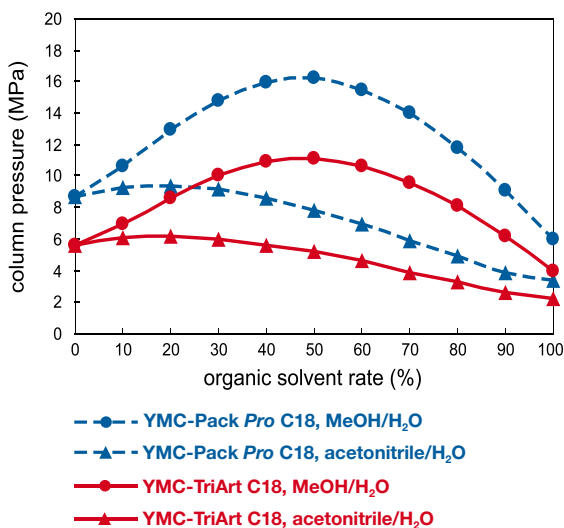


X-Bridge HILIC



The new uniform spherical particle support is used for YMC-TriArt C18 and C8. The particle is produced using **micro-reactor** technology for the granulation process. This results in reduction of the back-pressure and leads to more reproducibility in surface modification.

Low column back-pressure



Column: YMC-TriArt C18, 5 μm, 150 x 4.6 mm ID
 Part-No.: TA12S051546PT
 Eluent: acetonitrile / water or methanol / water
 Flow rate: 1.0 ml/min
 Temperature: 25 °C

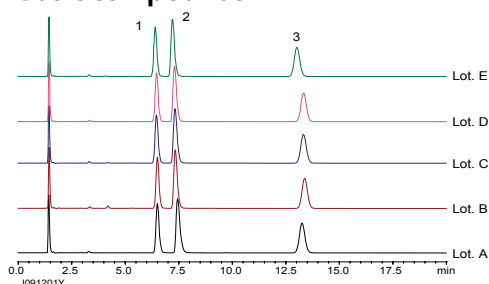
The revolutionary production technique, adapted from micro-reactor flow technology, produces a multi-layered silica/organic hybrid stationary phase, with outstanding narrow pore size and particle size distributions which result in low back pressures.

YMC-TriArt C18 is designed for use under many conditions. Elution with higher viscosity methanol (compared with acetonitrile), YMC-TriArt C18 generates lower pressure (approx 30% lower than with conventional phases).

Batch-to-Batch reproducibility

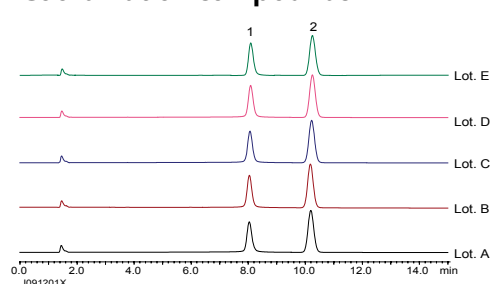
Excellent reproducibility of YMC-TriArt C18 and C8 is available even for the analysis for basic and coordination compounds which normally exhibit tailing and adsorption effects.

Basic compounds



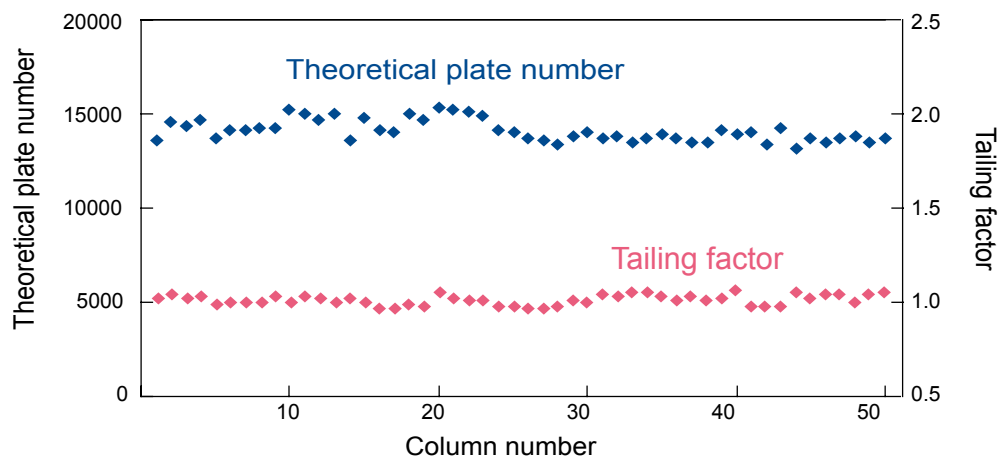
Column: YMC-TriArt C18, 5 μ m, 150 x 3.0 mm ID
 Eluent: 20 mM KH_2PO_4 (pH 6.9) / acetonitrile (65/35)
 Flow rate: 0.425 ml/min
 Temperature: 40 $^\circ\text{C}$
 Detection: UV at 235 nm

Coordination compounds



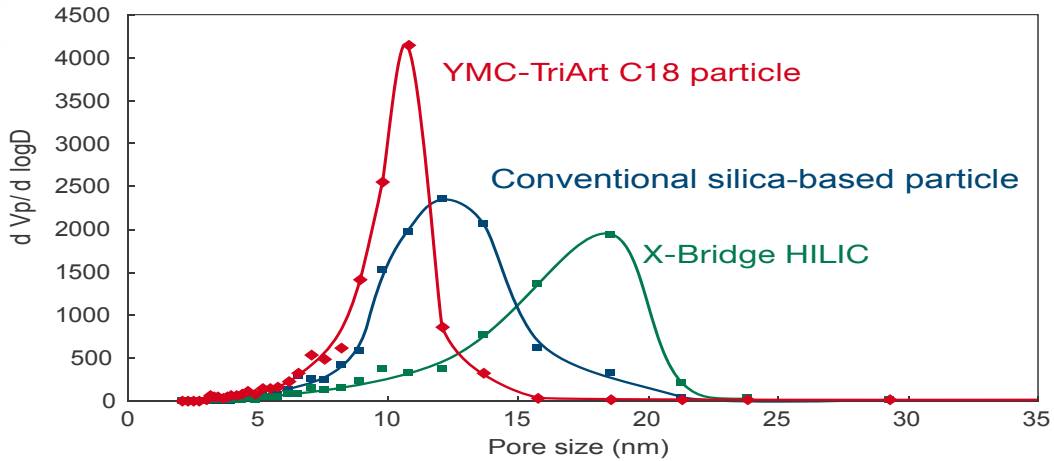
Column: YMC-TriArt C18, 5 μ m, 150 x 3.0 mm ID
 Eluent: acetonitrile / 0.1% H_3PO_4 (40/60)
 Flow rate: 0.425 ml/min
 Temperature: 40 $^\circ\text{C}$
 Detection: UV at 254 nm

The reproducibility of packed columns is shown below in terms of theoretical plate number (N) and tailing factor (Tf). YMC-TriArt C18 and C8 packed columns exhibit a very narrow range of variation.



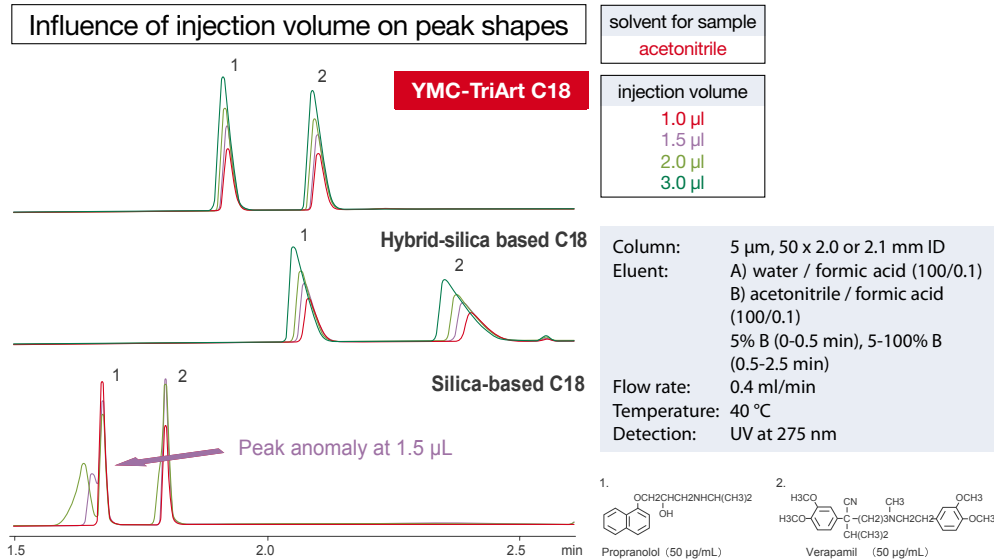
Column: YMC-TriArt C18, 5 μ m, 150 x 4.6 mm ID
 Part-No.: TA12S051546PT
 Eluent: acetonitrile / water (40/60)
 Flow rate: 1.0 ml/min
 Temperature: ambient
 Sample: butyl benzoate

Narrow pore distribution



This figure shows the pore size distributions of some competitive materials. Comparing the pore size distributions of some competitive materials shows that YMC-TriArt has a narrower distribution which results in sharper peak shapes.

Improved loadability

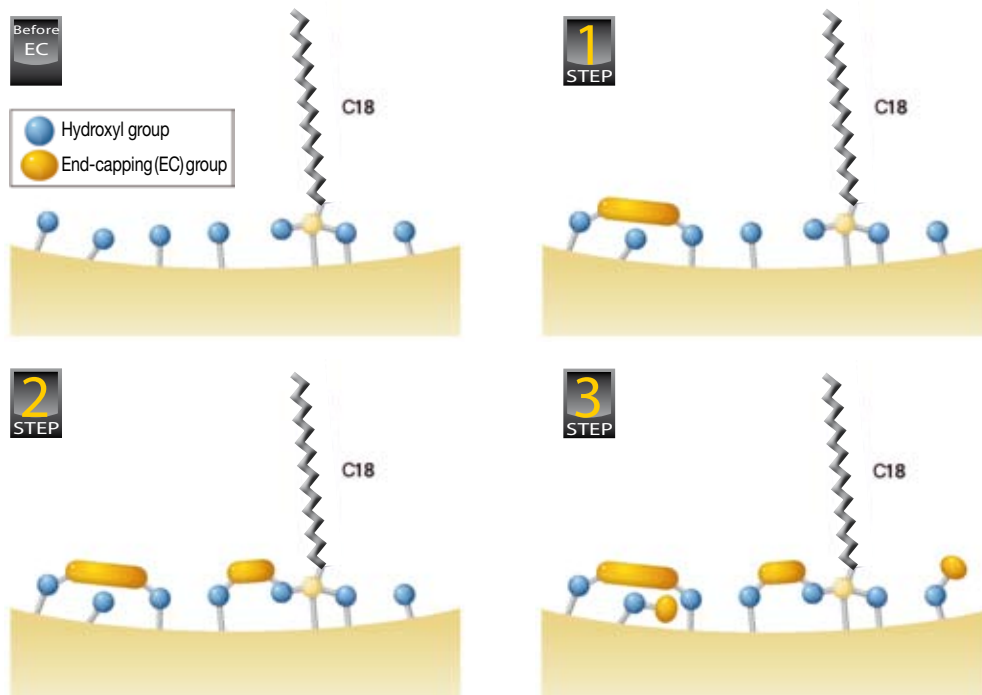


In order to prevent peak errors, there is the limit to the injection volume when the sample is injected in high elution solvents (such as 100% acetonitrile). Compared with traditional columns, more than double the injection volume can be injected into YMC-TriArt columns as a result of the extremely narrow particle size distribution.

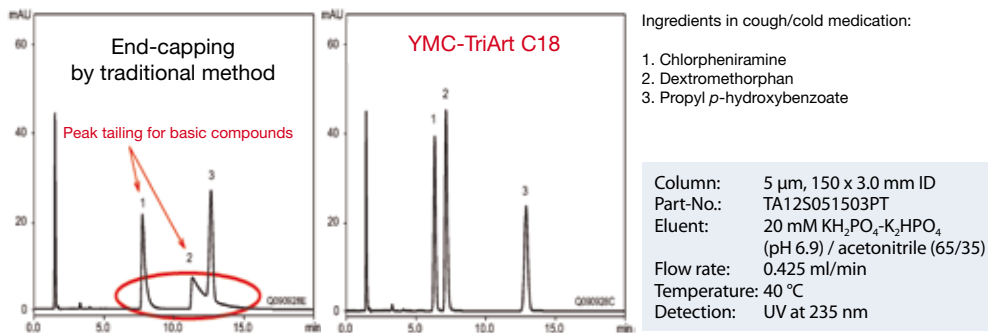
Multi-stage endcapping

After bonding the alkyl chain, there are highly reactive and less reactive silanols on the surface. In traditional bonding processes, these are reacted with a single capping-compound in one step. However, the highly reactive silanols can be hydrolysed easily which contributes to the poor durability. The less reactive silanols are hard to endcap which results in poor resolution due to peak tailing.

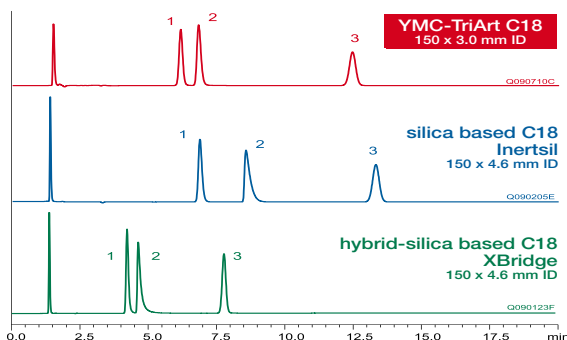
YMC-TriArt C18 and C8 use a new innovation in end capping called "multistage end-capping" for its surface modification process. By using a number of compounds with the different reactivities in successive steps, all silanols can be capped to the maximum extent.



The chromatographic result of a "good" end-capping is demonstrated:



Basic compounds



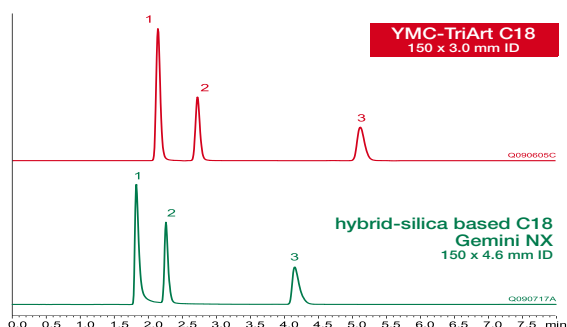
Ingredients in a cough/cold medication

1. Chlorpheniramine
2. Dextromethorphan
3. Propyl p-hydroxybenzoate

Column: 5 μ m, 150 x 3.0 or 150 x 4.6 mm ID
 Eluent: 20 mM KH_2PO_4 - K_2HPO_4 (pH 6.9) / acetonitrile (65/35)
 Flow rate: 0.425 ml/min for 3.0 mm ID
 1.0 ml/min for 4.6 mm ID
 Temperature: 40 °C
 Detection: UV at 235 nm

The innovative surface modification technology results in excellent peak shapes even for basic compounds that often exhibit peak tailing with conventional silica- and hybrid silica-based reversed phase columns.

Acidic compounds



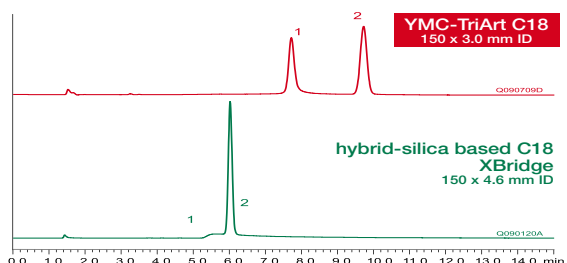
Organic acid

1. Formic acid
2. Acetic acid
3. Propionic acid

Column: 5 μ m, 150 x 3.0 or 150 x 4.6 mm ID
 Eluent: methanol / 0.1% H_3PO_4 (5/95)
 Flow rate: 0.425 ml/min for 3.0 mm ID
 1.0 ml/min for 4.6 mm ID
 Temperature: 37 °C
 Detection: UV at 210 nm

YMC-TriArt phases is synthesised using methodology adapted from micro-reactor technology. This technique ensures a reduction of impurities that contribute to peak tailing during the analysis of some types acidic compounds.

Coordinating compounds



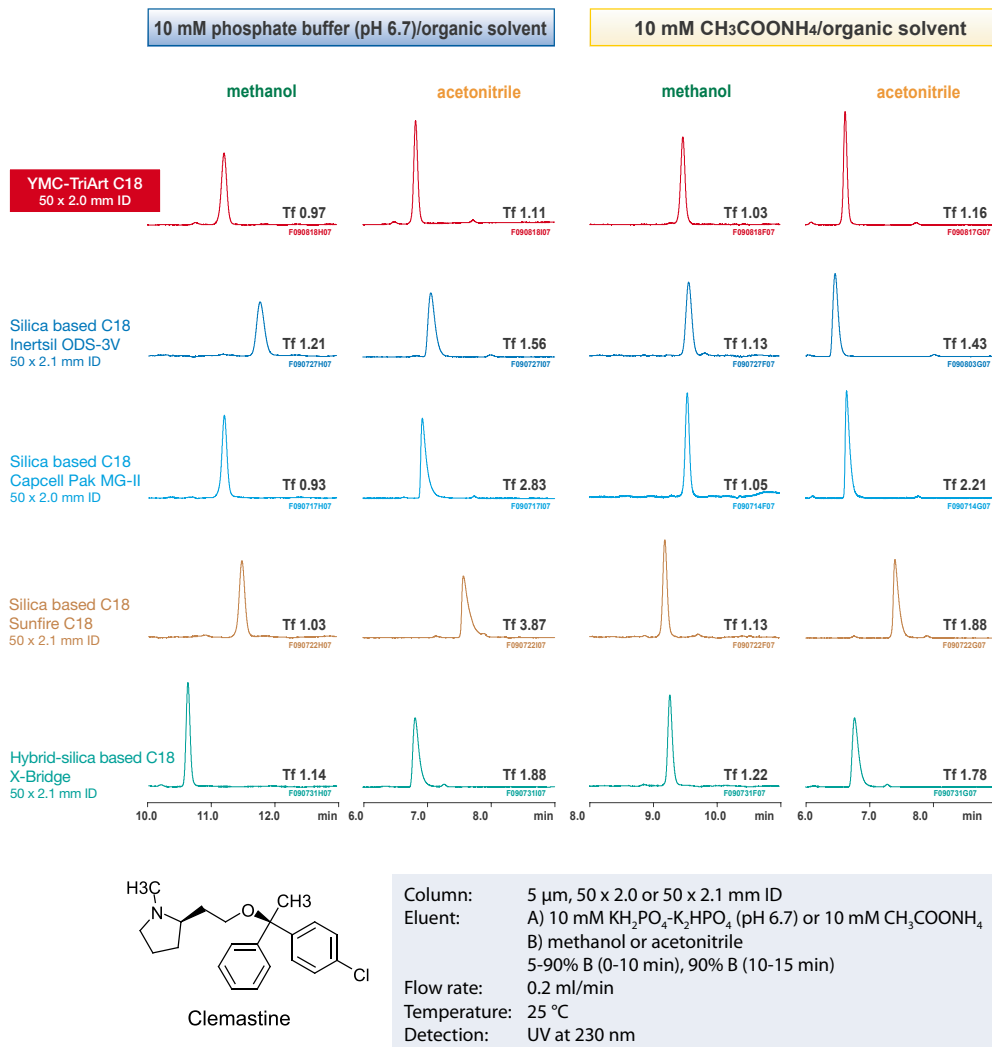
Hinokitiol

1. Hinokitiol
2. Methyl benzoate

Column: 5 μ m, 150 x 3.0 or 150 x 4.6 mm ID
 Eluent: acetonitrile / 0.1% H_3PO_4 (40/60)
 Flow rate: 0.425 ml/min for 3.0 mm ID
 1.0 ml/min for 4.6 mm ID
 Temperature: 40 °C
 Detection: UV at 254 nm

YMC-TriArt phases have an extremely low level of metal impurities, much lower than conventional products, ensuring excellent peak shape for coordination compounds.

Comparison of clemastine analysis



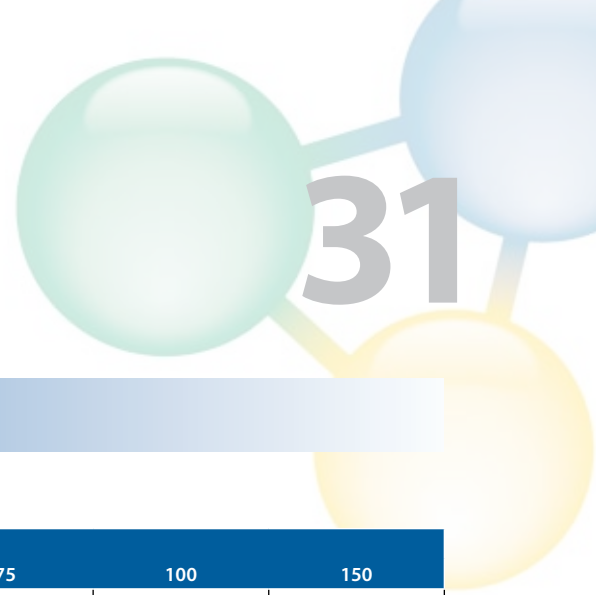
Clemastine is a well-known basic compound which readily exhibits peak tailing with conventional ODS columns. YMC-TriArt C18 provides sharp separations with many different buffer/solvent compositions.

Substance index

A	page	E	page	P	page
Acesulfam-K	9	Endorphins	23	Paraquat	11
Acetic acid	28	Enkephalins	23	Pentobarbital	15
6"-O-Acetylaidzin	21	Erythorbic acid	21	Peonidin	22
6"-O-Acetylgenistin	21	Erythromycin	10,16	Peonidin-3-O-arabinside	22
6"-O-Acetylglycitin	21	Erythromycin estolate	16	Peonidin-3-O-galactoside	22
Aciclovir	18	Erythromycin ethylsuccinate	16	Peonidin-3-O-glucoside	22
Agrochemicals	7	Etizolam	12	Peptides	23
Acidic compounds	28			Pesticides	7,14
Alkaloids	15	F		Petunidin	22
Amlodipine besilate	19,20	Flavonoids	17	Petunidin-3-O-arabinside	22
Angiotensin I	10	Flazasulfuron	7	Petunidin-3-O-galactoside	22
Angiotensin II	10	Formic acid	28	Petunidin-3-O-glucoside	22
Angiotensin III	10	Fungicides	7	Phenobarbital	15
Anthocyanidins	22			Pindolol	11
Anthocyanins	22	G		Procine insulin	10
Aromatic carboxylic acids	17	Genistein	21	Propranolol	11,26
Artificial sweeteners	9	Genistin	21	Propionic acid	28
L-Ascorbic acid	21	Glycitein	21	Propyl paraben	6,8
L-Ascorbic acid 2-glucoside	21	Glycitin	21	Propyl p-hydroxybenzoate	27,28
Asulam	7			Pyridoxine HCl	21
Atenolol	7	H			
Atorvastatin calcium hydrate	20	Halosulfuronmethyl	7	Q	
Atropine	15	Herbicides	7	Quinine	15
Azoxystrobin	7	Hexobarbital	15		
		Hinokitiol	28	R	
B		Human insulin	10	Riboflavin	21
Baicalein	17	Hydrochlorothiazide	20		
Barbital	15			S	
Barbiturates	15	I		Saccharin	9
Benzodiazepine derivatives	12	Insulin	10	Salicylic acid	17
Basic compounds	28	Isobutyl p-hydroxybenzoate	19	Scopolamine	15
Basic drugs	20			Secobarbital	15
Betablockers	11	K		Siduron	7
Bovine insulin	10	Kaempferol	17	Soy isoflavones	21
				Spriarmycin	10
C		L		Sulpha drugs	10
Candesartan cilexetil	20	Lincomycin HCl	19	Sulphamerazin	10
Carvedilol	20	Luteolin	17	Sulphamethaxole	10
Chlorphenamine	8			Sulphathiazole	10
Chlorpheniramine	6,27,28	M			
Chlortetracycline	11,13	Macrolide antibiotics	10	T	
trans-Cinnamic acid	17	6"-O-Malonyldaidzin	21	Tetracycline antibiotics	11,13
Cinchonine	15	6"-O-Malonylgenistin	21	Thiamine HCl	21
Clemastine	29	6"-O-Malonylglycitin	21	Thiram	7
Clinamycin HCl	19	Malvidin	22	Triazolam	12
Coordinating compounds	28	Malvidin-3-O-arabinside	22	Triclopyr	7
Copper 8-quinolinolate	7	Malvidin-3-O-galactoside	22		
Cyanidin	22	Malvidin-3-O-glucoside	22	U	
Cyanidin-3-O-arabinside	22	Mecoprop	7	Uracil	11
Cyanidin-3-O-galactoside	22	Methyl benzoate	28		
Cyanidin-3-O-glucoside	22	Methyl p-hydroxybenzoate	17	V	
Cyanocobalamin	21	Metoprolol	9	Valsartan	20
Cyclamat-Na	9			Verapamil	26
		N		Vitamin B1	21
D		Nadolol	11	Vitamin B2	21
Daidzein	17,21	Neurotensin	23	Vitamin B6	21
Daidzin	21	Nicotinamide	21	Vitamin B12	21
Delphinidin	22	Nicotinic acid	18,21	Vitamin C	21
Delphinidin-3-O-arabinside	22			W	
Delphinidin-3-O-galactoside	22	O		Water-soluble vitamins	21
Delphinidin-3-O-glucoside	22	Organic acids	28		
Dextromethorphan	6,8,27,28	Oxine-copper	7		
Dihydroquinine	15	Oxytetracycline	11,13		
Diquat	11	Oxytocin	23		
Doxycycline	13				

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Ordering Information

YMC-TriArt 1.9 µm UHPLC columns

Phase	Column ID (mm)	Column length (mm)					
		20	30	50	75	100	150
C18	2.0	TA12SP90202PT	TA12SP90302PT	TA12SP90502PT	TA12SP9L502PT	TA12SP91002PT	TA12SP91502PT
	3.0	—	TA12SP90303PT	TA12SP90503PT	TA12SP9L503PT	TA12SP91003PT	—
C8	2.0	TO12SP90202PT	TO12SP90302PT	TO12SP90502PT	TO12SP9L502PT	TO12SP91002PT	TO12SP91502PT
	3.0	—	TO12SP90303PT	TO12SP90503PT	TO12SP9L503PT	TO12SP91003PT	—

YMC-TriArt 3 µm analytical columns

Phase	Column ID (mm)	Column length (mm)					Guard cartridges with 10 mm length (pack of 5)
		50	75	100	150	250	
C18	2.0	TA12S030502WT	TA12S03L502WT	TA12S031002WT	TA12S031502WT	—	TA12S030102
	3.0	TA12S030503WT	TA12S03L503WT	TA12S031003WT	TA12S031503WT	—	TA12S030103
	4.6	TA12S030546WT	TA12S03L546WT	TA12S031046WT	TA12S031546WT	TA12S032546WT	TA12S030104
C8	2.0	TO12S030502WT	TO12S03L502WT	TO12S031002WT	TO12S031502WT	—	TO12S030102
	3.0	TO12S030503WT	TO12S03L503WT	TO12S031003WT	TO12S031503WT	—	TO12S030103
	4.6	TO12S030546WT	TO12S03L546WT	TO12S031046WT	TO12S031546WT	TO12S032546WT	TO12S030104

YMC-TriArt 5 µm analytical columns

Phase	Column ID (mm)	Column length (mm)					Guard cartridges with 10 mm length (pack of 5)
		50	75	100	150	250	
C18	2.0	TA12S050502WT	TA12S05L502WT	TA12S051002WT	TA12S051502WT	—	TA12S050102
	3.0	TA12S050503WT	TA12S05L503WT	TA12S051003WT	TA12S051503WT	—	TA12S050103
	4.6	TA12S050546WT	TA12S05L546WT	TA12S051046WT	TA12S051546WT	TA12S052546WT	TA12S050104
C8	2.0	TO12S050502WT	TO12S05L502WT	TO12S051002WT	TO12S051502WT	—	TO12S050102
	3.0	TO12S050503WT	TO12S05L503WT	TO12S051003WT	TO12S051503WT	—	TO12S050103
	4.6	TO12S050546WT	TO12S05L546WT	TO12S051046WT	TO12S051546WT	TO12S052546WT	TO12S050104

YMC-TriArt, 12 nm, 5 µm in ACTUS high-throughput semipreparative hardware

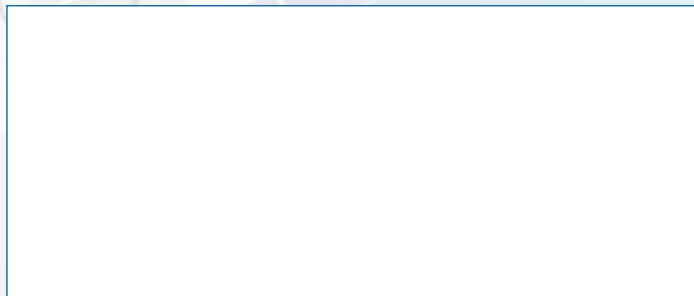
Phase	Column ID (mm)	Column length (mm)			
		50	75	100	150
C18	10.0	TA12S050510WX	—	TA12S051010WX	TA12S051510WX
	20.0	TA12S050520WX	—	TA12S051020WX	TA12S051520WX
	30.0	TA12S050530WX	TA12S05L530WX	TA12S051030WX	TA12S051530WX
C8	10.0	TO12S050510WX	—	TO12S051010WX	TO12S051510WX
	20.0	TO12S050520WX	—	TO12S051020WX	TO12S051520WX
	30.0	TO12S050530WX	TO12S05L530WX	TO12S051030WX	TO12S051530WX

YMC-TriArt, preparative bulk media, also available in 10 and 15 µm.

Available in pack sizes 100 mg, 500 mg, 1 kg, 5 kg



Your local distributor:



YMC Co., Ltd.

YMC Karasuma-Gojo Bld. 284 Daigo-cho,
Karasuma Nisuiru Gojo-dori Shimogyo-ku,
Kyoto 600-8106 Japan
TEL. +81(0)75-342-4515, FAX +81(0)75-342-4550
www.ymc.co.jp

YMC Europe GmbH

Schöttmannshof 19
D-46539 Dinslaken
Germany
TEL. +49(0)2064/427-0, FAX +49(0)2064/427-222
www.ymc.de

YMC America, Inc.

941 Marcon Boulevard Suite 301
Allentown, PA18109 USA
TEL. +1-610-266-8650, FAX +1-610-266-8652
www.ymc-america.com

YMC India Ltd.

CX - 07, 3rd Floor, Lobe - 1,
Tower - A, The Corenthum, Plot No- A-41,
Sector - 62, Noida - 201301 (UP) India.
TEL. +91(0)120-4276020 - 25, FAX +91(0)120-4276026
www.ymcindia.com

YMC Co., Ltd. Shanghai Rep. Office

Far East International Plaza A2404
No. 319 Xianxia Road, Shanghai 200051
P.R. China
TEL: +86-21-6235-1388, FAX: +86-21-6235-1398